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## EFFECTOR PROTEINS OF RAPAMYCIN

### Related Applications

5 This application is a continuation-in-part of co-pending patent application Serial No. 08/384,524, filed February 13, 1995, which is a continuation-in-part of patent application Serial No. 08/312,023, filed September 26, 1994, now abandoned, which is a continuation-in-part of patent application Serial No. 08/207,975, filed March 8, 1994, now abandoned.

10 This invention concerns effector proteins of Rapamycin. More particularly, this invention concerns novel Rapamycin-FKBP12 binding proteins of mammalian origin for identification, design and synthesis of immunomodulatory, anti-restenosis or anti-tumor agents.

15 BACKGROUND OF THE INVENTION

Rapamycin is a macrolide antibiotic produced by *Streptomyces hygroscopicus* which was first characterized via its properties as an antifungal agent. It adversely affects the growth of fungi such as *Candida albicans* and *Microsporum gypseum*. Rapamycin, its preparation and its antibiotic activity were described in U.S. Patent No. 3,929,992, issued December 30, 1975 to Surendra Sehgal et al. In 1977 Martel, R. R. et al. reported on immunosuppressive properties of rapamycin against experimental allergic encephalitis and adjuvant arthritis in the Canadian Journal of Physiological Pharmacology, 55, 48-51 (1977). In 1989, Calne, R. Y. et al. in Lancet, 1989, no. 2, p. 227 and Morris, R. E. and Meiser, B. M. in Medicinal Science Research, 1989, No. 17, P. 609-10, separately reported on the effectiveness of rapamycin in inhibiting rejection *in vivo* in allograft transplantation. Numerous articles have followed describing the immunosuppressive and rejection inhibiting properties of rapamycin, and clinical investigation has begun for the use of rapamycin in inhibiting rejection in transplantation in man.

35 Rapamycin alone (U.S. Patent 4,885,171) or in combination with picibanil (U.S. Patent 4,401,653) has been shown to have antitumor activity. R. R. Martel et al. [Can. J. Physiol. Pharmacol. 55, 48 (1977)] disclosed that rapamycin is effective in the experimental allergic encephalomyelitis model, a model for multiple sclerosis; in the

adjuvant arthritis model, a model for rheumatoid arthritis; and effectively inhibited the formation of IgE-like antibodies.

The immunosuppressive effects of rapamycin have been disclosed in FASEB 3, 5 3411 (1989). Cyclosporin A and FK-506, other macrocyclic molecules, also have been shown to be effective as immunosuppressive agents, therefore useful in preventing transplant rejection [FASEB 3, 3411 (1989); FASEB 3, 5256 (1989); R. Y. Calne et al., Lancet 1183 (1978); and U.S. Patent 5,100,899].

10 Rapamycin has also been shown to be useful in preventing or treating systemic lupus erythematosus [U.S. Patent 5,078,999], pulmonary inflammation [U.S. Patent 5,080,899], insulin dependent diabetes mellitus [Fifth Int. Conf. Inflamm. Res. Assoc. 121 (Abstract), (1990)], and smooth muscle cell proliferation and intimal thickening following vascular injury [Morris, R. J. Heart Lung Transplant 11 (pt. 2): 197 (1992)].

15 Mono- and diacylated derivatives of rapamycin (esterified at the 28 and 43 positions) have been shown to be useful as antifungal agents (U.S. Patent 4,316,885) and used to make water soluble prodrugs of rapamycin (U.S. Patent 4,650,803). Recently, the numbering convention for rapamycin has been changed; therefore 20 according to Chemical Abstracts nomenclature, the esters described above would be at the 31- and 42- positions. U.S. Patent 5,118,678 discloses carbamates of rapamycin that are useful as immunosuppressive, anti-inflammatory, antifungal, and antitumor agents. U.S. Patent 5,100,883 discloses fluorinated esters of rapamycin. U.S. Patent 5,118,677 discloses amide esters of rapamycin. U.S. Patent 5,130,307 discloses 25 aminoesters of rapamycin. U.S. Patent 5,117,203 discloses sulfonates and sulfamates of rapamycin. U.S. Patent 5,194,447 discloses sulfonylcarbamates of rapamycin.

30 U.S. Patent No. 5,100,899 (Calne) discloses methods of inhibiting transplant rejection in mammals using rapamycin and derivatives and prodrugs thereof. Other chemotherapeutic agents listed for use with rapamycin are azathioprine, corticosteroids, cyclosporin (and cyclosporin A), and FK-506, or any combination thereof.

Rapamycin produces immunosuppressive effects by blocking intracellular signal transduction. Rapamycin appears to interfere with a calcium independent

signalling cascade in T cells and mast cells [Schreiber et al. (1992) Tetrahedron 48:2545-2558]. Rapamycin has been shown to bind to certain immunophilins which are members of the FK-506 binding proteins (FKBP) family. In particular, Rapamycin has been shown to bind to the binding proteins, FKBP12, FKBP13, FKBP25 [Galat 5 A. et al., (1992) Biochemistry 31(8):2427-2437 and Ferrera A, et al., (1992) Gene 113(1):125-127; Armistead and Harding, Ann. Reports in Med. Chem. 28:207-215, 1993], and FKBP52 [WO 93/07269]

Rapamycin is able to inhibit mitogen-induced T cell and B cell proliferation as 10 well as proliferation induced by several cytokines, including IL-2, IL-3, IL-4 and IL-6 (reviewed by Sehgal et al., Med. Research Rev.14: 1-22, 1994). It can also inhibit antibody production. Rapamycin has been shown to block the cytokine-induced activation of p70<sup>S6</sup> kinase which appears to correlate with Rapamycin's ability to decrease protein synthesis accompanying cell cycle progression (Calvo et al., Proc. 15 Natl. Acad. Sci. USA, 89:7571-7575,1992; Chung et al., Cell 69:1227-1236, 1992; Kuo et al., Nature 358:70-73,1992; Price et al., Science 257:973-977, 1992). It also inhibits the activation of cdk2/cyclin E complex (Flanagan et al., Ann. N.Y.Acad. Sci, in press; Flanagan et al, Mol. Cell biol., in press; Flanagan et al., J.Cell Biochem. 17A:292, 1993). Rapamycin's effects are not mediated by direct binding to p70<sup>S6</sup> 20 kinase and cdk2/cyclin E, but by action of the Rapamycin-FKBP complex on upstream component(s) which regulate the activation status of the kinases.

It is generally accepted that the action of immunosuppressive drugs, such as Rapamycin, cyclosporine and FK506, is dependent upon the formation of a complex 25 with their respective intracellular receptor proteins called immunophilins. While the binding of these immunosuppressants with their respective immunophilins inhibits the cis-trans peptidyl prolyl isomerase (PPIase) activity of immunophilins, PPIase inhibition is not sufficient to mediate the immunosuppressive activity (reviewed in Armistead and Harding, Annual Reports in Med. Chem, 28:207-215:1993). Two 30 rapamycin analogs which are Diels Alder adducts, one with 4-phenyl-1,2,4-triazoline-3,5-dione, and the second with 4-methyl-1,2,4-triazoline-3,5-dione, bind to FKBP, inhibited its PPIase activity, yet they did not exhibit any detectable immunosuppressive activity. The phenyl-triazolinedione Diels Alder adduct at high molar excess has been shown to competitively inhibit rapamycin's effect on DNA synthesis in mitogen-

stimulated murine thymocyte proliferation (Ocain et al., Biochem. Biophys. Res. Commun. 192:1340, 1993). Recent evidence suggests that the binary immunophilin-drug complex such as cyclophilin-cyclosporin A and FKBP-FK506 gains a new function that enables it to block signal transduction by acting on specific target proteins.

5 The molecular target of both cyclophilin-cyclosporin A and FKBP-FK506 complexes such as has been identified as the  $\text{Ca}^{+2}$ /calmodulin dependent serine/threonine phosphatase calcineurin (J. Liu et al, Cell 66, 807, 1991; J. Liu et al, Biochemistry 31, 3896, 1992; W.M. Flanagan, et al., Nature 352, 803, 1992; McCaffrey et al., J. Biol. Chem. 268, 3747, 1993; McCaffrey et al., Science 262:750, 1993).

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Rapamycin's antifungal and immunosuppressive activities are mediated via a complex consisting of Rapamycin, a member of the FK506 binding protein (FKBP) family and at least one additional third protein, called the target of Rapamycin (TOR). The family of FKBPs is reviewed by Armistead and Harding (Annual Reports in Med. Chem, 28:207-215:1993). The relevant FKP molecule in Rapamycin's antifungal activity has been shown to be FKP12 (Heitman et al., Science 253:905-909:1993). In mammalian cells, the relevant FKBPs are being investigated. Although two TOR proteins (TOR1 and TOR2) have been identified in yeast (Kunz et al., Cell 73:585-596:1993), the target of Rapamycin in human cells remains elusive. The carboxy terminus of yeast TOR2 has been shown to exhibit 20% identity with two proteins, the p110 subunit of phosphatidylinositol 3-kinase and VPS34, a yeast vacuolar sorting protein also shown to have PI 3K activity. However, J. Blenis et al. (AAI meeting, May, 1993) have reported that Rapamycin-FKP12 complex does not directly mediate its effects on PDGF stimulated cells via the p110, p85 PI 3K complex.

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#### DESCRIPTION OF THE INVENTION

This invention concerns isolated, cloned and expressed proteins which bind to a complex of GST-FKP12-Rapamycin. These proteins are isolated from membrane preparations of Molt 4 T cell leukemia. The sizes of the four novel proteins are estimated by PAGE migration to be  $125\pm12$  kilodaltons (kDa),  $148\pm14$  kDa,  $208\pm15$  kDa and  $210\pm20$ kDa and will be referred to herein and in the claims that follow, as the 125 kDa, 148 kDa, 208 kDa, and 210 kDa, respectively. The four proteins may also be referred to herein as effector proteins.

The proteins of this invention can be used in screening assays, such as enzyme inhibitor assays and binding assays, to identify endogenous complexes and ligands and novel exogenous compounds (like Rapamycin) which modulate their functions. They 5 can also be used in assays to identify compounds with therapeutic benefit for restenosis, immunomodulation and as antitumor agents. Cloning the proteins of this invention does not only allow the production of large quantities of the proteins, it also provides a basis for the development of related anti-sense therapeutics. The use of 10 cDNA clones to generate anti-sense therapeutics with immunomodulatory activity (for use against transplantation rejection, graft versus host disease, autoimmune diseases such as lupus, myasthenia gravis, multiple sclerosis, rheumatoid arthritis, type I diabetes, and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, pulmonary inflammation, asthma, and eye uveitis), antirestenosis and anti-tumor activity is included within the scope of this invention.

15       The proteins of the present invention can be isolated from mammalian cells, such as cells of the T cell leukemia cell line, Molt 4 (ATCC 1582, American Type Cell Culture, 12301 Parklawn Drive, Rockville, MD, USA, 20852), the B cell lymphoma, BJAB, or normal human T cells. These mammalian cells can be lysed in a buffer 20 containing protease inhibitors and reducing agent (2-ME), such as hypotonic buffer A (100 mM HEPES, pH 7.5, 20 mM KCl, 1 mM EDTA, 0.4 mM PMSF and 2 mM beta mercaptoethanol (2-ME)). The cell nuclei and unbroken cells are cleared by centrifugation at a temperature which minimizes protein degradation. The membrane fraction of the cells can then be concentrated or pelleted by ultracentrifugation at 25 100,000 g. Detergent solubilization of the membrane pellet is carried out in a detergent containing buffer such as buffer B (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM KCl, 0.2 mM PMSF, 1 mM 2-ME, 2 mM CaCl<sub>2</sub>, 2 mM MgCl<sub>2</sub>, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), containing CHAPSO (3-[(3-cholamido-propyl)dimethylammonio]-1-propane sulfonate; 12 mM) or Triton X100 (polyethylene 30 glycol 4-isoctylphenyl ether). The solubilized membrane proteins can then be separated from the debris by 100,000g ultracentrifugation at a temperature which minimizes protein degradation. The supernatant containing solubilized membrane proteins is then preabsorbed with an affinity resin, such as glutathione resin, in the presence of protease inhibitors at a temperature which minimizes protein degradation..

After centrifugation to remove the resin from the supernatant, the supernatant is then incubated with complexed Rapamycin or Rapamycin analog to FKBP, such as GST-FKBP12--Rapamycin at a temperature which minimizes protein degradation. The mixture of solubilized membrane proteins, incubated with complexed Rapamycin or 5 Rapamycin analog to FKBP, such as GST-FKBP12--Rapamycin, can then be incubated with the affinity resin to bind the complexes of rapamycin or rapamycin analog, FKBP fusion protein and binding proteins at a temperature which minimizes protein degradation. After most non-specific proteins are rinsed away using a detergent containing buffer, such as Buffer C (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM 10 KCl, 0.2 mM PMSF, 1 mM 2-ME or 10 mM dithiothreitol, 0.5 mM CaCl<sub>2</sub>, 0.5 mM MgCl<sub>2</sub>, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain and 0.1% Triton X100) (Polyethylene glycol 4-isooctyl phenyl ether), the proteins are eluted from the resin under denaturing conditions, such as a buffer containing sufficient detergent to 15 dissociate it from resin (e.g. Laemmli buffer with or without glycerol or dye, as described by Laemmli, Nature 227:680, 1970), or non-denaturing conditions such as a buffer containing an appropriate eluting compound for the affinity column, such as 5 mM glutathione. The proteins can then be separated by size using SDS polyacrylamide gel electrophoresis (SDS-PAGE).

20 The present invention also includes the genomic DNA sequences for the abovementioned proteins, as well as the cDNA and anti-sense RNA and DNA sequences which correspond to the genes for the abovementioned proteins. The present invention further includes the proteins of other mammalian species which are homologous or equivalent at least in function to the abovementioned proteins, as well 25 as the DNA gene sequences for the homologous or equivalent proteins and the cDNA and anti-sense RNA and DNA sequences which correspond to the genes for the homologous or equivalent proteins.

For the purposes of this disclosure and the claims that follow, equivalents of the 30 proteins of this invention are considered to be proteins, protein fragments and/or truncated forms with substantially similar, but not identical, amino acid sequences to the proteins mentioned above, the equivalents exhibiting rapamycin-FKBP complex binding characteristics and function similar to the proteins mentioned above. Therefore, in this specification and the claims below, references to the 125 kDa, 148

kDa, 208 kDa, and 210 kDa proteins of this invention are also to be understood to indicate and encompass homologous or equivalent proteins, as well as fragmented and/or truncated forms with substantially similar, but not identical, amino acid sequences of the 125 kDa, 148 kDa, 208 kDa, and 210 kDa proteins mentioned above.

5        These proteins or protein homologues or equivalents can be generated by similar isolation procedures from different cell types and/or by recombinant DNA methods and may be modified by techniques including site directed mutagenesis. For example, the genes of this invention can be engineered to express one or all of the 10 proteins as a fusion protein with the fusion partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc, glutathione S-transferase, FLAG etc). Mutations or truncations which result in a soluble form can also be generated by site directed mutagenesis and would give advantages in isolation.

15      This invention further includes oligopeptide fragments, truncated forms and protein fragments that retain binding affinity yet have less than the active protein's amino acid sequences. This invention also includes monoclonal and polyclonal antibodies specific for the proteins and their uses. Such uses include methods for screening for novel agents for immunomodulation and/or anti-tumor activity and 20 methods of measuring the parent compound and/or metabolites in biological samples obtained from individuals taking immunosuppressive drugs. The use of the cDNA clone to generate anti-sense therapeutics (Milligan et al, J. Med. Chem. 36:1923-1936, 1993) with immunomodulatory activity (transplantation rejection, graft versus host disease, autoimmune diseases such as lupus, myasthenia gravis, multiple sclerosis, 25 rheumatoid arthritis, type I diabetes, and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, pulmonary inflammation, asthma, and eye uveitis), and anti-tumor activity is also included in the present invention.

30      The proteins of this invention can also be made by recombinant DNA techniques familiar to those skilled in the art. That is, the gene of the protein in question can be cloned by obtaining a partial amino acid sequence by digestion of the protein with a protease, such as Lysine C, and isolating the resulting protein fragments by microbore HPLC, followed by fragment sequencing (Matsudaira in A Practical

Guide to Protein and Peptide Purification for Microsequencing, Academic Press (San Diego, CA, 1989)). The determined sequence can then be used to make oligonucleotide probes which can be used to screen a human cDNA library directly or generate probes by polymerase chain reaction. The library can be generated from 5 human T cells or the cell lines, Molt 4, Jurkat, or other etc. to obtain clones. These clones can be used to identify additional clones containing additional sequences until the protein's full gene, i.e. complete open reading frame, is cloned.

It is known in the art that some proteins can be encoded by an open reading 10 frame which is longer than initially predicted by the size of the protein. These proteins may represent cleavage products of the precursor protein translated from the complete open reading frame (eg. IL-1 beta) or proteins translated using a downstream start codon (eg. Hepatitis B surface antigen). In view of this knowledge, it is understood that the term cDNA as used herein and in the claims below refers to cDNA for the 15 gene's complete open reading frame or any portions thereof which may code for a protein of this invention or the protein's fragments, together or separate, or truncated forms, as previously discussed.

In a complementary strategy, the gene(s) for the proteins of this invention may 20 be identified by interactive yeast cloning techniques using FKBP12:RAPA as a trap for cloning. These strategies can also be combined to quicken the identification of the clones.

The relevant cDNA clone encoding the gene for any of the four proteins can 25 also be expressed in E. coli, yeast, or baculovirus infected cells or mammalian cells using state of the art expression vectors. Isolation can be performed as above or the cDNA can be made as a fusion protein with the fusion partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc, glutathione S-transferase, etc). Mutations which result in a soluble form can also be generated by site directed 30 mutagenesis and would give advantages in isolation.

The uses of such cDNA clones include production of recombinant proteins. Further, such recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, or fragments thereof (including peptide oligomers) are useful in

generation of antibodies to these proteins. Briefly, monoclonal or polyclonal antibodies are induced by immunization with recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, or fragments thereof (including peptide oligomers conjugated to a carrier protein (e.g. keyhole limpet hemocyanin or bovine serum albumin)) of animals using state of the art techniques. The antibodies can be used in the purification process of the natural proteins isolated from mammalian cells or recombinant proteins from *E. coli*, yeast, or baculovirus infected cells or mammalian cells, or cell products.

10       The uses of such cDNA clones include production of recombinant proteins. Further, such recombinant proteins, or the corresponding natural proteins isolated from mammalian cells, are useful in methods of screening for novel agents such as synthetic compounds, natural products, exogenous or endogenous substrates for immunomodulation and/or antitumor activity. The natural products which may be screened may include, but are not limited to, cell lysates, cell supernatants, plant extracts and the natural broths of fungi or bacteria. As an example of a competitive binding assay, one of these proteins attached to a matrix (either covalently or noncovalently) can be incubated with a buffer containing the compounds, natural products, cell lysates or cell supernatants and a labeled rapamycin:FKBP complex. The ability of the compound, natural products, exogenous or endogenous substrates to competitively inhibit the binding of the complex or specific antibody can be assessed. Examples of methods for labeling the complex include radiolabeling, fluorescent or chemiluminescent tags, fusion proteins with FKBP such as luciferase, and conjugation to enzymes such as horse radish peroxidase, alkaline phosphatase, acetylcholine esterase (ACHE), etc. As an example of an enzymatic assay, the proteins are incubated in the presence or absence of novel agents such as synthetic compounds, natural products, exogenous or endogenous substrates with substrate and the enzymatic activity of the protein can be assessed. Methods of measuring the parent compound and/or metabolites in biological samples obtained from individuals taking immunosuppressive drugs can also be assessed using these proteins.

30       This invention includes a method for identifying substances which may be useful as immunomodulatory agents or anti-tumor agents, the method utilizing the following steps:

- 10 -

a) combining the substance to be tested with one of the four mammalian proteins (125 kDa, 148kDa, 208 kDa or 210 kDa) of this invention, with the protein being bound to a solid support:

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b) maintaining the substance to be tested and the protein bound to the solid support of step (a) under conditions appropriate for binding of the substance to be tested with the protein, and

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c) determining whether binding of the substance to be tested occurred in step (b).

This invention also includes a method for identifying substances which may be useful as immunomodulatory or anti-tumor agents which involves the following steps:

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a) combining a substance to be tested with one of the mammalian proteins of this invention, the protein being bound to a solid support:

20

b) maintaining the substance to be tested and the protein bound to the solid support of step (a) under conditions appropriate for binding of the substance to be tested with the protein, and

c) determining whether the presence of the substance to be tested modulated the activity of the mammalian protein.

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This invention further includes a method for detecting, in a biological sample, rapamycin, rapamycin analogs or rapamycin metabolites which, when complexed with a FKBP, bind to one of the four listed proteins of this invention, the method comprising the steps of:

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a) combining the biological sample with a FKBP to form a first mixture containing, if rapamycin, rapamycin analogs or rapamycin metabolites are present in the biological sample, a rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes;

- b) creating a second mixture by adding the first mixture to one of the proteins of this invention, the protein bound to a solid support ;
- 5 c) maintaining the second mixture of step (b) under conditions appropriate for binding the rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes, if present, to the protein of this invention; and
- 10 d) determining whether binding of the rapamycin:FKBP complexes, rapamycin analog:FKBP complexes, or rapamycin metabolite:FKBP complexes and the protein occurred in step (c).

Also included in this invention is the use of the cDNA clones to generate anti-sense therapeutics. This can be accomplished by using state of the art techniques, such as those described in Milligan et al, J. Med. Chem. 36:14:1924-1936. For the purposes of this disclosure and the claims that follow, antisense RNA and DNA are understood to include those RNA and DNA strands derived from a cDNA clone which encodes for one of the four proteins (125 kDa, 148 kDa, 208 kDa or 210 kDa) of the present invention which have a native backbone or those which utilize a modified backbone. Such modifications of the RNA and DNA backbones are described in Milligan et al, J. Med. Chem. 36:14:1924-1936. The antisense compounds created by the state of the art techniques recently described (Milligan et al, J. Med. Chem. 36:14:1924-1936) can be useful in modulating the immune response and thus useful in the treatment or inhibition of transplantation rejection such as kidney, heart, liver, lung, bone marrow, pancreas (islet cells), cornea, small bowel, and skin allografts, and heart valve xenografts; in the treatment or inhibition of autoimmune diseases such as lupus, rheumatoid arthritis, diabetes mellitus, myasthenia gravis, and multiple sclerosis; and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, and eye uveitis. The antisense molecules of this invention can have antitumor, antifungal activities, and antiproliferative activities. The compounds of this invention therefore can be also useful in treating solid tumors, adult T-cell leukemia/lymphoma, fungal infections, and hyperproliferative vascular diseases such as restenosis and atherosclerosis. Thus, the present invention also comprises

methods for treating the abovementioned maladies and conditions in mammals, preferably in humans. The method comprises administering to a mammal in need thereof an effective amount of the relevant antisense therapeutic agent of this invention.

5        When administered for the treatment or inhibition of the above disease states, the antisense molecules of this invention can be administered to a mammal orally, parenterally, intranasally, intrabronchially, transdermally, topically, intravaginally, or rectally.

10      It is contemplated that when the antisense molecules of this invention are used as an immunosuppressive or antiinflammatory agent, they can be administered in conjunction with one or more other immunoregulatory agents. Such other immunoregulatory agents include, but are not limited to azathioprine, corticosteroids, such as prednisone and methylprednisolone, cyclophosphamide, rapamycin, 15     cyclosporin A, FK-506, OKT-3, and ATG. By combining the complexes of this invention with such other drugs or agents for inducing immunosuppression or treating inflammatory conditions, the lesser amounts of each of the agents are required to achieve the desired effect. The basis for such combination therapy was established by Stepkowski whose results showed that the use of a combination of rapamycin and 20     cyclosporin A at subtherapeutic doses significantly prolonged heart allograft survival time. [Transplantation Proc. 23: 507 (1991)].

25      Treatment with these antisense compounds will generally be initiated with small dosages less than the optimum dose of the compound. Thereafter the dosage is increased until the optimum effect under the circumstances is reached. Precise dosages will be determined by the administering physician based on experience with the individual subject treated. In general, the antisense compounds of this invention are most desirably administered at a concentration that will afford effective results without causing any harmful or deleterious side effects.

30      In light of the therapeutic value of the abovementioned antisense compounds, this invention also includes pharmaceutical compositions containing the antisense RNA and antisense DNA compounds derived from cDNA clones which encode for the 125 kDa, 148 kDa, 208 kDa and 210 kDa proteins of this invention.

This invention also comprises the following process for isolating the proteins of this invention, as well as the proteins isolated therefrom:

5        A process for isolating proteins from mammalian cells, the process comprising the steps of:

10        1.        The mammalian cells of interest are grown and harvested. As mentioned previously, the cells may be of T cell origin (e.g. T cell lymphomas, leukemias, normal human T cells), B cell origin (e.g. EBV transformed B cells, normal human B cells), mast cells, or other cell sources sensitive to rapamycin. The cells may be processed shortly after harvesting or may be stored frozen, such as in pellets, prior to processing. The cells which are kept frozen may be stored in a dry ice and ethanol bath, stored frozen at -70-80° C until use. This step of growing and harvesting the 15 cells of interest may be seen as the first step of this process or as merely preparatory for the present process.

20        2.        Cells are lysed in a buffer containing a buffering agent (e.g. HEPES, Tris, pH 7.5), low salt (e.g. 10 -50 mM NaCl or KCl), chelating agent (e.g. 1-2 mM EDTA), protease inhibitors (e.g. 0.4 mM PMSF) and a reducing agent (e.g. 2 mM 2-ME or 1-20 mM Dithiothreitol) at a temperature which minimizes protein degradation (e.g. 4 °C). It should be understood that the mammalian cells may be treated in any manner capable of producing cell lysis, including sonic lysis and douncing.

25        3.        Unbroken cells and cell nuclei are precleared from lysates by centrifugation at a temperature which minimizes protein degradation (e.g. 4 °C). Centrifugation at, for example, 1600g for 10 minutes has been found sufficient to preclear the unbroken cells and cell nuclei from the lysates. This step, while not 30 mandatory, provides a clearer preparation for the steps that follow.

4.        The membrane fraction in the precleared lysate is then concentrated, such as by ultracentrifugation. An example of this concentration would be ultracentrifugation at 100,000 g for 1-1.5 hours.

5. The membrane proteins (e.g. transmembrane, integral and membrane associated proteins) are then solubilized. This may be accomplished by incubating the pellet of Step 4 in a buffer containing a detergent which solubilizes the proteins without detrimentally denaturing them, a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 1 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl<sub>2</sub>, 0-5 mM MgCl<sub>2</sub>) at a temperature which minimizes protein degradation (e.g. 4 °C). Examples of detergents useful in this step are CHAPSO (3-[3-cholamidopropyl]dimethylammonio]-1-propane sulfonate) or Triton X100 (polyethylene glycol 4-isooctylphenyl ether). After this step, the mixture contains solubilized membrane proteins and non-solubilized cellular debris.

15 6. The solubilized membrane proteins are separated from the non-solubilized cellular debris, such as by ultracentrifugation (eg 100,000g for 1-1.5 hours) at a temperature which minimizes protein degradation (e.g. 4 °C).

20 7. The supernatant containing solubilized membrane proteins is incubated with an affinity resin in a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 10 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl<sub>2</sub>, 0-5 mM MgCl<sub>2</sub>) at a temperature and time which allows the absorption of the proteins which bind to affinity resin directly, and minimizes protein degradation (e.g. 4 °C).

30 8. The resin is then removed from the supernatant by centrifugation at a temperature which minimizes protein degradation (e.g. 4 °C).

9. The supernatant is then incubated with Rapamycin or Rapamycin analog (IC50 in LAF < 500nM) complexed to fusion protein of FKBP12 +protein which enhances the isolation of the desired effector protein and through which the fusion protein binds to an affinity resin or affinity column, such as GST-FKBP12,

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Histidine oligomer -FKBP12, biotin-FKBP12, etc., in a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salt (e.g. 100 - 200 mM NaCl + 20 mM KCl), reducing agent (e.g. 1-2 mM 2-ME or 1 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain),  
5 divalent cations (e.g. 0-5 mM CaCl<sub>2</sub>, 0-5 mM MgCl<sub>2</sub>) at a temperature and for a time which allows binding of the effector proteins to the fusion FKBP protein:Rapamycin or analog complexes and minimizes protein degradation (e.g. 4 °C and 1-2 hours).

10. The mixture of Step 9 containing the effector proteins and fusion FKBP protein:Rapamycin complexes is incubated with an affinity resin at a temperature and for a time which allows binding of the complexes of the effector proteins and fusion FKBP protein:Rapamycin or analog to the affinity resin and minimizes protein degradation (e.g. 4 °C and 0.5-2 hours).

15. Most non-specific proteins are rinsed away from the resin using a buffer which dissociates binding of non-specific proteins but not the complex between the desired proteins and RAPA-FKBP, such as a buffer containing a buffering agent (e.g. 20-50 mM Tris or HEPES, pH 7.2), salts (e.g. 100 - 1000 mM NaCl, KCl), reducing agent (e.g. 1-2 mM 2-ME or 10 - 20 mM dithiothreitol), protease inhibitors (e.g. 0.2 mM PMSF, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), divalent cations (e.g. 0-5 mM CaCl<sub>2</sub>, 0-5 mM MgCl<sub>2</sub>) and detergent which dissociates binding of non-specific proteins but not the complex between the four proteins and RAPA-fusion FKBP protein such as Triton X100 (Polyethylene glycol 4-isooctyl phenyl ether).

25. The effector proteins and the fusion FKBP protein:Rapamycin complexes are eluted from the resin using an appropriate buffer, such as a buffer containing sufficient detergent to dissociate it from resin (e.g. Laemli buffer with or without glycerol or dye, Laemli, Nature 227:680, 1970), or an appropriate eluting compound for the affinity column, such as glutathione, histidine.  
30

13. The effector proteins can then be separated by size. This may be accomplished in any manner which separates the proteins by size, including, but not

limited to, polyacrylamide gel electrophoresis and size exclusion column chromatography.

It might also be useful to compare the proteins isolated by a control procedure,  
5 that is a procedure which substitutes buffer for the rapamycin or rapamycin analog with  
an IC<sub>50</sub> in LAF < 500 nM in step 8, can be used to more easily distinguish proteins  
which bind to the rapamycin:FKBP complex.

The proteins of this invention can also be made by recombinant DNA  
10 techniques familiar to those skilled in the art. That is, the gene of the protein in  
question can be cloned by obtaining a partial amino acid sequence by digestion of the  
protein with an appropriate endopeptidase, such as Lysine C, and isolating the  
resulting protein fragments by microbore HPLC, followed by fragment sequencing  
(Matsudaira in A Practical Guide to Protein and Peptide Purification for  
15 Microsequencing, Academic Press, San Diego, CA 1989). The determined sequence  
can then be used to make oligonucleotide probes which can be used to screen a human  
cDNA library, such as those for human T cells, Molt 4, Jurkat, etc, to obtain  
clones.(Sambrook, Fritsch, and Maniatis, Molecular Cloning, A Laboratory Manual,  
Cold Spring Harbor Press, 1989) These clones can be used to identify additional  
20 clones containing additional sequences until the protein's full gene is cloned  
(Sambrook, Fritsch, and Maniatis, Molecular Cloning, A Laboratory Manual, Cold  
Spring Harbor Press, 1989). In a complementary strategy, the gene(s) may be  
identified by interactive yeast cloning techniques using FKBP12:RAPA as a trap for  
cloning (Chien et al., Proc. Natl. Acad. Sci. 88: 9578-9582, 1991). These strategies  
25 can also be combined to quicken the identification of the clones.

The relevant cDNA clone can also be expressed in E.coli, yeast, or baculovirus  
infected cells or mammalian cells using state of the art expression vectors. Isolation can  
be performed as above or the cDNA can be made as a fusion protein with the fusion  
30 partner giving an advantage in isolation (e.g. HIS oligomer, immunoglobulin Fc,  
glutathione S-transferase, etc). Mutations which result in a soluble form can also be  
generated by site directed mutagenesis and would give advantages in isolation.

Homologs in the mouse, rat, monkey, dog and other mammalian species can be obtained using similar procedures. In addition, upon isolation of the human clone of the proteins, the clone can be used to screen for homologs in other mammalian species. These homologs can also be used to develop binding assays and to set up high throughput screening assays for compounds, endogenous ligands, exogenous ligands with immunomodulatory activity.

Compounds, endogenous ligands and exogenous ligands having such immunomodulatory activity would be useful in modulating the immune response and thus useful in the treatment or inhibition of transplantation rejection such as kidney, heart, liver, lung, bone marrow, pancreas (islet cells), cornea, small bowel, and skin allografts, and heart valve xenografts; in the treatment or inhibition of autoimmune diseases such as lupus, rheumatoid arthritis, diabetes mellitus, myasthenia gravis, and multiple sclerosis; and diseases of inflammation such as psoriasis, dermatitis, eczema, seborrhea, inflammatory bowel disease, and eye uveitis.

The compounds, endogenous ligands and exogenous ligands mentioned above can also have antitumor, antifungal activities, and antiproliferative activities. The compounds of this invention therefore can be also useful in treating solid tumors, adult T-cell leukemia/lymphoma, fungal infections, and hyperproliferative vascular diseases such as restenosis and atherosclerosis.

#### EXAMPLE 1

The proteins of the present invention were isolated utilizing a fusion protein of glutathione S-transferase--FK506 binding protein12 (GST-FKBP). GST-FKBP is produced by a recombinant E. coli containing the plasmid, pGEX-FKBP. The cells were grown, induced with IPTG and the fusion protein was isolated using standard technology described in D.B. Smith and K.S. Johnson, Gene 67, 31, 1988 and K.L. Guan and J.E. Dixon, Anal. Biochem. 192, 262, 1991. The solution containing glutathione and GST-FKBP12 was exchanged 5x using a Centricon 10 filtration unit (Amicon) to remove the glutathione and exchange the buffer.

Molt 4 cells ( $1 \times 10^9$ ) were grown in standard media (RPMI 1640 containing 100 U/ml penicillin, 100 ug/ml L-glutamine, 10% FCS). The cells were harvested and rinsed 3x with PBS (50mM phosphate buffer, pH 7.0, 150 mM NaCl), flash frozen in dry-ice ethanol bath and stored at -80°C. On ice, the cells were thawed and lysed using 5 a dounce homogenizer with B pestle in 5 ml of buffer A (10 mM Hepes, pH 7.5, 20 mM KCl, 1 mM EDTA, 0.4 mM PMSF and 2 mM 2-ME). After the debris was cleared by centrifugation at 1600g for 10 min. and the membrane fraction was concentrated by 100,000g centrifugation (1 hour), the 100,000 g pellet was incubated in 3 ml buffer B (50 mM Tris, pH 7.2, 100 mM NaCl, 20 mM KCl, 0.2 mM PMSF, 10 1 mM 2-ME, 2 mM CaCl<sub>2</sub>, 2 mM MgCl<sub>2</sub>, 5 µg/ml aprotinin, leupeptin, pepstatin A and antipain), containing 12 mM CHAPSO for two hours at 4°C. The solubilized membrane proteins were separated from the debris by a 100,000 g centrifugation. After preabsorption of the supernatant for 3-18 hours with 0.4 ml glutathione sepharose 15 resin swollen in buffer B, the supernatant was incubated with complexed Rapamycin-GST-FKBP12 (preformed by incubation of 660 ug GST-FKBP + 60 ug RAPA in buffer B for 1-2 hours, 4°C) for two hours at 4°C. The supernatant was then incubated for 2 hours at 4°C with 100 ul glutathione resin (1:1 Buffer B). Nonspecific proteins were rinsed 5x with buffer C (buffer B + 0.1% Triton x 100) and the proteins eluted from the resin in Laemli buffer by incubation at 95°C for 3 minutes and 20 microcentrifugation. The proteins were separated by size using a 7% SDS-PAGE followed by silver stain. Four bands corresponding to proteins of molecular weights of 210kDa, 208 kDa, 148 kDa, and 125 kDa were present in higher concentrations in the sample containing RAPA + GST-FKBP12 vs GST-FKBP alone.

25 The mitogen-stimulated thymocyte proliferation assay called the LAF, can be inhibited by rapamycin or analogs such as demethoxyrapamycin and indicates relative activity of rapamycin analogs in immunosuppression. The same proteins were isolated using GST-FKBP complexed with the immunosuppressive analog, demethoxyrapamycin (Table 1). The Diels Alder adducts bound to FKBP12 and 30 inhibited PPIase activity of FKBP12 but did not exhibit detectable immunosuppressive activity and thus do not bind to the target of rapamycin. The use of these two compounds complexed with GST-FKBP12 in the analogous isolation procedure (ie. replacing rapamycin:GST-FKBP12) yielded background levels of the 210kDa proteins (no rapamycin)(Table 1). FK506, is an immunosuppressive compound which binds to

- 19 -

FKBP and mediates at least some of its effects through the binding of the FK506-FKBP complex with calcineurin. FK506 when complexed with GST-FKBP in an analogous procedure yielded only background levels of the 210 kDa protein (Table 1).

5

TABLE 1  
Comparison of Binding of Rapamycin  
Analog--FKBP12 complexes to 210 kDa Protein

	<u>Compound</u>	<u>210 kDa</u>	<u>LAF</u>	<u>PPIase(Ki)</u>
10	RAPA	+++	6 nM	0.12nM
	demethoxyrapamycin	+++	58nM	4.4 nM
15	Diels Alder adduct (phenyl)	±	>1000nM	12 nM
	Diels Alder adduct (methyl)	±	>1000nM	12 nM
	FK506	±	3nM*	0.4 nM
	none (FKBP)	±		

(\* mechanism of action is different)

20

It is known that rapamycin must bind to a member of the FKBP family in order to mediate its effects. To verify that the proteins of this invention bind to the complex RAPA-GST-FKBP and not individually to rapamycin or FKBP12, a modified isolation procedure was employed. The modification consists of using (1) a rapamycin-42-biotin glycinate ester in place of rapamycin (both exhibit equivalent immunosuppressive activity in the LAF assay), (2) no exogenous FKBP and (3) a streptavidin-conjugated resin in place of glutathione-resin. Only background levels of the 210 kDa protein was isolated using this modified isolation procedure.

25

The 210 kDa protein was isolated using the GST--FKBP12--rapamycin complex from BJAB cells (B cell lymphoma) and normal human T lymphocytes purified by Ficoll-Hypaque and T cell columns.

- 20 -

The results of the partial amino acid composition analysis are set forth in Table 2, below. It should be noted that the percentage of the basic amino acids was not determined.

5

TABLE 2

	Peak Number	Component Name	Retention Time	Peak Area	Response Factor	Peak Height	Concentration No./50ul
10			9.38				
			11.09				
	1	Asp/Asn	12.06	12.47076	0.02344	0.05142	0.30
	2	Thr	13.05	2.92898	0.00000	0.00985	0.068
	3	Ser	13.78	6.43968	0.00000	0.01995	0.15
15			15.68				
	4	Glu/Gln	16.87	25.47273	0.00000	0.05285	0.59
		Prp	18.24				0.14
	5	Gly	22.35	21.50384	0.00000	0.04645	0.44
			22.90				
20	6	Ala	23.73	16.69160	0.00000	0.03113	0.36
			26.06				
			28.81				
	7	Val	29.39	4.83196	0.00000	0.00605	0.11
		Met	32.28				
25	8	Ile	34.10	3.00560	0.2326	0.00782	0.0699
	9	Leu	35.09	5.73202	0.02331	0.01372	0.1383
	10	nLeu	36.27	20.48232	0.02174	0.04286	0.4453

TABLE 2 (Cont'd)

Peak Number	Component Name	Retention Time	Peak Area	Response Factor	Peak Height	Concentration No./50μl
5	11	Tyr	38.33	1.44792	0.02618	0.00226
	12	Phe	40.05	1.25017	0.02703	0.00187
	13	His	47.79	1.50905	0.02553	0.00580
	14		51.80	12.66136	0.00000	0.0000
10	15	Lys	53.34	9.90767	0.02283	0.2262
<hr/>				146.53645	0.33436	<hr/>
15	Not Determined			144.29		

EXAMPLE 2

The 210 kDa ( $210 \pm 20$  kDa) protein of this invention was isolated from 4 x 20  $10^{11}$  Molt 4 cells using the affinity matrix protocol as described previously. Bound proteins were eluted from the affinity matrix with 1x Laemli buffer without glycerol and dye (0.0625 M Tris-HCl, pH 6.8, 2% SDS, 0.37M b-mercaptoethanol) and were concentrated 3 consecutive times by centrifugation using centricon 100 (Amicon, Beverly, MA) at 4 °C the first two times and at 18 °C the third time. The concentrated 25 sample was eluted from the centricon 100 filter by incubating 2 hours at room temperature with an equal volume of 2 x laemli buffer without glycerol and dye the first 2 x and 2 x laemli buffer the third time. The proteins in the sample were separated by PAGE on a 1.5mm thick 7% polyacrylamide gel (38:1). The proteins were transferred to polyvinylidene difluoride, PVDF, (Biorad, Hercules, CA) in 10 x Tris/glycine buffer 30 (Biorad) containing 0.037% SDS at 50 mAmps at 4 °C overnight. The proteins on the PVDF were stained with amido black (Biorad) in 10% ethanol, 2% acetic acid and the appropriate band was excised, rinsed with PBS and water and stored frozen.

Sequencing

The protein (approx. 3 ug) on the PVDF membrane was digested in situ with trypsin using a modification described by J. Fernandez et al, (Anal.Biochem. 201: 255-64, 1992 ). Briefly, the PVDF was cut into 1 mm<sup>2</sup> pieces, prewet, and the protein digested in a 100mM Tris-HCl, pH buffer containing 10% acetonitrile, and 1% reduced triton (CalBiochem) with 0.2ug trypsin at 37 °C for 6 hours followed by addition of 0.2 ug trypsin and incubation overnight. The fragments were eluted from the membrane by sonication and the buffer containing the fragments were separated by microfuge centrifugation. The membranes were backextracted 2x (i.e., 50 ul buffer was added to membranes, sonicated, and centrifuged in a microfuge and solution pooled with the original buffer containing the eluted fragments.) The sample (140-145 ul) was separated by narrow bore high performance liquid chromatography using a Vydac C18 2.1mm x 150 mm reverse phase column on a Hewlett Packard HPLC 1090 with a 40 diode array detector as described previously by W.Lane et al, (J.Protein Chem., 10(2): 151-60, 1991). Multiple fractions were collected and measured for absorption at multiple wavelengths (210, 277 and 292 nm). Optimal fractions were chosen for sequencing based on resolution, symmetry, and ultraviolet absorption and spectra (210 nm, 277 nm and 292 nm).. An aliquot (5%) of the optimal fractions was analyzed for homogeneity and length of fragment by matrix assisted laser desorption time of flight mass spectrometry, MALDE-TOF-MS, on a Finnigan lasermat. Selected optimal fractions were sequenced by automated Edman degradation on an Applied Biosystems 477A protein sequencer using microcartridge and manufacturer's recommended chemistry cycle.

25

Sequence comparison

Comparison was performed using the Intelligenetics suite (Intelligenetics, CA) .  
30

Sequences

Utilizing the methods mentioned above, it was determined that the 210 kDa (210±20 kDa) protein of this invention contains peptide fragments, four of which have amino acid sequences as shown below:

- a) ILLNIEHR;
- B) LIRPYMEPILK;
- 5 c) DXMEAQE; and
- d) QLDHPLPTVHPQVTYAYM(K)

Those skilled in the art will recognize the one-letter symbols for the amino acids  
10 in question (the definitions for which can also be seen at page 21 of the text  
*Biochemistry*, Third Edition, W.H. Freeman and Company, © 1988 by Lubert Stryer). Those so skilled will also understand that the X in sequence c) indicates an as yet unidentified amino acid and the parentheses in sequence d) indicates that the amino acid  
in the position in question is possibly lysine.

15 As mentioned previously, the present invention includes fragmented or truncated forms of the proteins mentioned herein. This includes proteins which have as part or all of their amino acid sequence one or more of the four sequences listed as a)-d), above. For the purposes of the claims, below, the proteins referred to as including  
20 one or more of the "internal amino acid sequences" are understood to be any protein which contains one of the sequences listed above, whether the protein is comprised wholly of one or more of the sequences a)-d) or whether one or more of the sequences mentioned above form any portion of the protein. This is understood to include all locations on the protein's amino acid sequence including, but not limited to, those  
25 sections of the protein which initiate and terminate the protein's amino acid chain.

These partial amino acid sequences were compared with sequences in the Genbank database. There was identity with the sequence, accession number L34075 (Brown et al., Nature 369, 756-758 (1994)). The cDNA of the SEP gene was cloned  
30 as follows: Two micrograms of Molt 4 cDNA (Clontech, Palo Alto, CA) in 1 x PCR buffer (10 mM Tris-HCl, pH 8.3, 50 mM KCl, 1 mM MgCl<sub>2</sub>, 200 μM dATP, 200 μM dTTP, 200 μM dCTP, 200 μM dGTP; Perkin Elmer, ) with 1 unit Taq polymerase (Perkin Elmer), was amplified by Polymerase chain reaction (PCR) at 94 C for 30 sec., 66 C for 4 min for 30 cycles, 72 C for 10 min by three separate reactions containing  
35 one of the following pairs of oligomers:

CGATCGGTCGACTGCAGCACTTGGGGATTGTGCTCTC and  
GCGGCCGCAAGCTTCTCATGCATGACAAACAGCCCAGGC; or  
GCGGCCGCAAGCTTCAAGTATGCAAGCCTGTGCGGCAAGA and  
CGATCGGTCGACACACCTCTGCATCAGAGTCAAGTGGTCA; or  
5 CGGGCCGCAAGCTCCTCAGCTCACATCCTAGAGCTGCA and  
CGATCGGTCGACTTATTACCAGAAAGGGACCAGCCAATATA .

The oligonucleotides were synthesized and isolated by methods previously described and known in the art (Chemical and Enzymatic Synthesis of Gene 10 Fragments, ed. by H.G.Gassin and Anne Lang, Verlag Chemie, FLA, 1982). The resulting PCR products named SEP3, SEP4, and SEP5, respectively, were incubated at 15°C overnight in buffer containing T4 DNA ligase (1 unit) and 50 ng pcII which was modified to efficiently ligate PCR products (TA cloning kit, Invitrogen, San 15 Diego, CA) to yield PCR-pcII ligated products. The PCR-pcII products were transformed into competent *E. coli* INValphaF cells obtained commercially from Invitrogen. Miniprep DNA was prepared using the Quiagen miniprep kits (Quiagen, Chatsworth, CA) and the clones containing the appropriate sized PCR product were identified by restriction enzyme digestion with commercially available HindIII or Sal I , electrophoresis, and comparison to standards. Sep2 and Sep1 cDNA was made using 20 the TimeSaver cDNA synthesis Kit (Pharmacia, Piscataway, NJ) with the first strand synthesis reaction containing oligodT (0.13 µg) and 250 pmoles of

CGATCGGTCGACCAGATGAGCACATCATAGCGCTGATGA or  
CGATCGGTCGACAAATTCAAAGCTGCCAAGCGTTCGGAG,  
25 respectively. Sep2 and Sep1 second strand synthesis was performed using the TimeSaver cDNA synthesis kit with the addition of 250 pmoles of

GCGGCCGCAAGCTTGGCTCGAGCAATGGGCCAGGCA or  
30 GCGGCCGCAAGCTTAAGATGCTTGGAACCGCACCTGCCG,  
respectively. The Sep2 and Sep1 cDNA was then amplified by PCR using

CGATCGGTCGACCAAGATGAGCACATCATAGCGCTGATGA and  
GCGGCCGCAAGCTTGGCTCGAGCAATGGGGCCAGGCA or  
GCGGCCGCAAGCTTAAGATGCTTGGAACCGCACCTGCCG and  
CGATCGGTCGACAAATTCAAAGCTGCCAAGCGTTGGAG,

19.  
20  
21  
22

5. respectively as described above. The Sep2 PCR products were cloned into the TA cloning kit (Invitrogen). The Sep 1 PCR products were digested with Hind III and Sal I, separated from the pCII vector by agarose electrophoresis. The Sep1 (HindIII-SalI) fragment was isolated using the Sepaglas bandprep kit from Pharmacia and cloned into the HindIII and Sal I sites of pUC19 as described (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989). Ligation of the isolated Sep2(HindIII, AspI) and Sep3(AspI, SalI) fragments or Sep4(HindIII, AccIII/MroI) and Sep5(AccIII/MroI, Sal I) fragments into pUC18(HindIII, SalI) vector and transformation of competent E. coli INValphaF cells (Invitrogen) was performed by techniques known to those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989) to obtain pUC18-Sep 23 and pUC18-Sep45 which contain nucleotides 1468- 5326 and 4964 - 7653, respectively, of the full length clone shown in the attached Sequence No. 1. Ligation of the pUC19-Sep1 (EcoRV, SalI), Sep2345 (EcoRV, SalI) fragments and transformation of competent E. coli INValphaF cells (Invitrogen) were performed by techniques known to those skilled in the art (as described by Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989) to obtain the full length clone. The nucleic acid sequence coding for this protein and its amino acid sequence are shown in Sequence No. 1.
- 10 25 A fusion protein, called glutathione S transferase-sirolimus effector protein, GST-SEP, was engineered by subcloning the Sep4 and Sep5 fragments into the plasmid, pGEX-KG (Guan, K. and Dixon, J.E. (1991) Anal. Biochem. 192, 262-267) as follows. Briefly, Sep4 was digested with commercially available HindIII restriction enzyme, the restriction site was filled in with the Klenow fragment of DNA polymerase 30 (Gibco), and the DNA was extracted with phenol-chloroform and ethanol precipitated using techniques known by those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989). The SEP4 (HindIII-Klenow) was further digested with MroI restriction enzyme, separated from the pCII vector by agarose electrophoresis and isolated as the fragment SEP4-HindIII-Klenow-MroI. Sep5 fragment was prepared by

digestion with SalI and MroI, separated from the p<sub>cII</sub> vector by agarose electrophoresis and isolated as the fragment SEP5-SalI-MroI. pGEX-KG (Guan, K. and Dixon, J.E. (1991) Anal. Biochem. 192, 262-267) was digested with Nco I, filled in with the Klenow fragment of DNA polymerase and the DNA was extracted with phenol-  
5 chloroform and ethanol precipitated, using techniques of those skilled in the art (Sambrook et al., Molecular Cloning Cold Spring Harbor, 1989). pGEX-KG (NcoI, Klenow) was further digested with Sal I, separated from the undigested vector by agarose electrophoresis and isolated as the vector pGEX-KG-NcoI-Klenow-SalI, using techniques of those skilled in the art. Ligation of the vector, pGEX-KG-NcoI-Klenow-  
10 SalI and Sep 4 (HindIII, MroI) and Sep5 (MroI, SalI) fragments and transformation into *E. coli* strain INValphaF cells (Invitrogen) using techniques of those skilled in the art yielded the plasmid, pGEX-Sep45. Other *E. coli* hosts such as BL21 can also be used. The DNA and protein sequence of this fusion protein is shown in Sequence No. 2.

15 Flag sequences and kinase recognition domain of heart muscle kinase can be added at the amino terminal end, by methods known in the art (see Chen et al., *Gene* 1994 Feb. 11; 139 (1): 73-75) within SEP or at the carboxy terminus of SEP, SEP4,5 or other fragments using an oligonucleotide which includes the coding sequence for Asp Tyr Lys Asp Asp Asp Lys. The fusion protein can be isolated by affinity chromatography with anti-flag specific antibodies using the commercially available kits from IBI, New Haven, Conn.

20 Transformed host cells containing sequences of this invention have been deposited with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, USA, and have been given the ATCC designations listed below:

	<u>Sequence</u>	<u>ATCC Designation</u>
30	a) pUC19-Sep1(nucleotides 1- 1785 of Sequence No. 1)	ATCC 69756
	b) pUC18-Sep23 (nucleotides 1468- 5326 of Sequence No. 1)	ATCC 69753

- c) pUC18-Sep45 (nucleotides 4964 - 7653 of Sequence No. 1) ATCC 69754
- 5 d) pUC19-Sep1-5 (ATCC 69756 1-7653 of sequence 1) ATCC 69829
- e) pGEX-Sep45 plasmids (Sequence 2 ) ATCC 69755.

10

EXAMPLE 3

The 210 kDa protein of this invention was also isolated by the techniques described in Example 1 utilizing the following rapamycin analogs:

- 15 a) 42-Deoxy-42-[1-(1,1-dimethylethoxy)-2-oxoethoxy] rapamycin (which is described in U.S. Pat. No. 5,233,036);
- b) 42-[O-[(1,1-Dimethylethyl)dimethylsilyl]] rapamycin (described in U.S. Pat. No. 5,120,842);
- 20 c) Rapamycin 42-ester with N-[1,1-dimethylethoxy]carbonyl]-N-methylglycine (described in U.S. Pat. No. 5,130,307);
- d) Rapamycin 42-ester with 5-(1,1-dimethylethoxy)-2-[[[1,1-dimethylethoxy]carbonyl]amino]-5-oxopentanoic acid ethyl acetate solvate three quarter hydrate (see U.S. Pat. No. 5,130,307);
- 25 e) Rapamycin 42-ester with N-[(1,1-dimethylethoxy)carbonyl]glycylglycine hydrate (see U.S. Pat. No. 5,130,307); and
- f) Rapamycin 42-ester with N<sub>2</sub>, N<sub>6</sub>-bis[(1,1-dimethylethoxy)carbonyl]-L-lysine (see U.S. Pat. No. 5,130,307).

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5

(i) APPLICANT: Molnar-Kimber, Katherine L.  
Failli, Amedeo F.  
Caggiano, Thomas J.  
Nakanishi, Koji  
10 Chen, Yanqiu

10

(ii) TITLE OF INVENTION: Effector Proteins of  
Rapamycin

15

(iii) NUMBER OF SEQUENCES: 2

(iv) CORRESPONDENCE ADDRESS:

20

(A) ADDRESSEE: Ronald W. Alice,  
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(B) STREET: 5 Giralda Farms  
(C) CITY: Madison  
(D) STATE: New Jersey  
(E) COUNTRY: USA  
(F) ZIP: 07940-0874

25

(v) COMPUTER READABLE FORM:

30

(A) MEDIUM TYPE: Diskette, 3.50 inch, 1.4 Mb storage  
(B) COMPUTER: Apple Macintosh  
(C) OPERATING SYSTEM: Macintosh 7.1  
(D) SOFTWARE: Microsoft Word

35

(vi) CURRENT APPLICATION DATA:

40

(A) APPLICATION NUMBER:  
(B) FILING DATE:  
(C) CLASSIFICATION:

45

(vii) PRIOR APPLICATION DATA:

50

(A) APPLICATION NUMBER: US 08/312,023  
(B) FILING DATE: 26-SEPTEMBER-1994  
(C) APPLICATION NO: US 08/207,975  
(E) FILING DATE: 08-MARCH-1994

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55

(A) NAME: Eck, Steven R.  
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(C) REFERENCE/DOCKET NUMBER: AHP-93167-2-C2

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (610) 902-2628  
(B) TELEFAX: (610) 688-0273

(2) INFORMATION FOR SEQ. ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 7653  
5 (B) TYPE: nucleic acid  
(C) STRANDEDNESS: double-stranded  
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: sense orientation of double-stranded cDNA to mRNA

(iii) HYPOTHETICAL: no

15 (iv) ANTISENSE: no

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Molt 4 human T-cell leukemia  
cells  
20 (B) STRAIN: ATCC Strain CRL 1582

(xi) SEQUENCE DESCRIPTION: SEQ. ID NO: 1

AAG ATG CTT GGA ACC GGA CCT GCC GCC ACC ACC GCT GCC ACC ACA 48  
Met Leu Gly Thr Gly Pro Ala Ala Ala Thr Thr Ala Ala Thr Thr  
25 1 5 10 15

TCT AGC AAT GTG AGC GTC CTG CAG CAG TTT GCC AGT GGC CTA AAG AGC 96  
Ser Ser Asn Val Ser Val Leu Gln Gln Phe Ala Ser Gly Leu Lys Ser  
20 25 30

30 CGG AAT GAG GAA ACC AGG GCC AAA GCC GCC AAG GAG CTC CAG CAC TAT 144  
Arg Asn Glu Glu Thr Arg Ala Lys Ala Ala Lys Glu Leu Gln His Tyr  
35 40 45

35 GTC ACC ATG GAA CTC CGA GAG ATG AGT CAA GAG GAG TCT ACT CGC TTC 192  
Val Thr MET Glu Leu Arg Glu MET Ser Gin Glu Glu Ser Thr Arg Phe  
50 55 60

40 TAT GAC CAA CTG AAC CAT CAC ATT TTT GAA TTG GTT TCC AGC TCA GAT 240  
Tyr Asp Gln Leu Asn His His Ile Phe Glu Leu Val Ser Ser Ser Asp  
65 70 75

45 GCC AAT GAG AGG AAA GGT GGC ATC TTG GCC ATA GCT AGC CTC ATA GGA 288  
Ala Asn Glu Arg Lys Gly Gly Ile Leu Ala Ile Ala Ser Leu Ile Gly  
80 85 90 95

50 GTG GAA GGT GGG AAT GCC ACC CGA ATT GGC AGA TTT GCC AAC TAT CTT 336  
Val Glu Gly Gly Asn Ala Thr Arg Ile Gly Arg Phe Ala Asn Tyr Leu  
100 105 110

CGG AAC CTC CTC CCC TCC AAT GAC CCA GTT GTC ATG GAA ATG GCA TCC 384  
Arg Asn Leu Leu Pro Ser Asn Asp Pro Val Val MET Glu MET Ala Ser  
115 120 125

- 30 -

AAG GCC ATT GGC CGT CTT GCC ATG GCA GGG GAC ACT TTT ACC GCT GAG 432  
Lys Ala Ile Gly Arg Leu Ala MET Ala Gly Asp Thr Phe Thr Ala Glu  
130 135 140

5 TAC GTG GAA TTT GAG GTG AAG CGA GCC CTG GAA TGG CTG GGT GCT GAC 480  
Tyr Val Glu Phe Glu Val Lys Arg Ala Leu Glu Trp Leu Gly Ala Asp  
145 150 155

10 CGC AAT GAG GGC CGG AGA CAT GCA GCT GTC CTG GTT CTC CGT GAG CTG 528  
Arg Asn Glu Gly Arg Arg His Ala Ala Val Leu Val Leu Arg Glu Leu  
160 165 170 175

15 GCC ATC AGC GTC CCT ACC TTC TTC CAG CAA GTG CAA CCC TTC TTT 576  
Ala Ile Ser Val Pro Thr Phe Phe Gln Gln Val Gln Pro Phe Phe  
180 185 190

20 GAC AAC ATT TTT GTG GCC GTG TGG GAC CCC AAA CAG GCC ATC CGT GAG 624  
Asp Asn Ile Phe Val Ala Val Trp Asp Pro Lys Gln Ala Ile Arg Glu  
195 200 205

25 GGA GCT GTA GCC GCC CTT CGT GCC TGT CTG ATT CTC ACA ACC CAG CGT 672  
Gly Ala Val Ala Ala Leu Arg Ala Cys Leu Ile Leu Thr Thr Gln Arg  
210 215 220

30 GAG CCG AAG GAG ATG CAG AAG CCT CAG TGG TAC AGG CAC ACA TTT GAA 720  
Glu Pro Lys Glu MET Gln Lys Pro Gln Trp Tyr Arg His Thr Phe Glu  
225 230 235

35 GAA GCA GAG AAG GGA TTT GAT GAG ACC TTG GCC AAA GAG AAG GGC ATG 768  
Glu Ala Glu Lys Gly Phe Asp Glu Thr Leu Ala Lys Glu Lys Gly MET  
240 245 250 255

40 AAT CGG GAT GAT CGG ATC CAT GGA GCC TTG TTG ATC CTT AAC GAG CTG 816  
Asn Arg Asp Asp Arg Ile His Gly Ala Leu Leu Ile Leu Asn Glu Leu  
260 265 270

45 GTC CGA ATC AGC ATG GAG GGA GAG CGT CTG AGA GAA GAA ATG GAA 864  
Val Arg Ile Ser Ser MET Glu Gly Glu Arg Leu Arg Glu Glu MET Glu  
275 280 285

50 GAA ATC ACA CAG CAG CTG GTA CAC GAC AAG TAC TGC AAA GAT CTC 912  
Glu Ile Thr Gln Gln Leu His Asp Lys Tyr Cys Lys Asp Leu  
290 295 300

55 ATG GGC TTC GGA ACA AAA CCT CGT CAC ATT ACC CCC TTC ACC AGT TTC 960  
MET Gly Phe Gly Thr Lys Pro Arg His Ile Thr Pro Phe Thr Ser Phe  
305 310 315

50 CAG GCT GTA CAG CCC CAG TCA AAT GCC TTG GTG GGG CTG CTG GGG 1008  
Gln Ala Val Gln Pro Gln Gln Ser Asn Ala Leu Val Gly Leu Leu Gly  
320 325 330 335

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TAC AGC TCT CAC CAA GGC CTC ATG GGA TTT GGG ACC TCC CCC AGT CCA 1056  
Tyr Ser Ser His Gln Gly Leu MET Gly Phe Gly Thr Ser Pro Ser Pro  
340 345 350  
5 GCT AAG TCC ACC CTG GTG GAG AGC CGG TGT TGC AGA GAC TTG ATG GAG 1104  
Ala Lys Ser Thr Leu Val Glu Ser Arg Cys Cys Arg Asp Leu MET Glu  
355 360 365  
10 GAG AAA TTT GAT CAG GTG TGC CAG TGG GTG CTG AAA TGC AGG AAT AGC 1152  
Glu Lys Phe Asp Gln Val Cys Gln Trp Val Leu Lys Cys Arg Asn Ser  
370 375 380  
15 AAG AAC TCG CTG ATC CAA ATG ACA ATC CTT AAT TTG TTG CCC CGC TTG 1200  
Lys Asn Ser Leu Ile Gln MET Thr Ile Leu Asn Leu Leu Pro Arg Leu  
385 390 395  
20 GCT GCA TTC CGA CCT TCT GCC TTC ACA GAT ACC CAG TAT CTC CAA GAT 1248  
Ala Ala Phe Arg Pro Ser Ala Phe Thr Asp Thr Gln Tyr Leu Gln Asp  
400 405 410  
25 ACC ATG AAC CAT GCC CTA AGC TGT GTC AAG AAG GAG AAG GAA CGT ACA 1296  
Thr MET Asn His Ala Leu Ser Cys Val Lys Lys Glu Lys Glu Arg Thr  
420 425 430  
30 GCG GCC TTC CAA GCC CTG GGG CTA CTT TCT GTG GCT GTG AGG TCT GAG 1344  
Ala Ala Phe Gln Ala Leu Gly Leu Leu Ser Val Ala Val Arg Ser Glu  
435 440 445  
35 CCC CCA AAG GAC TTC GCC CAT AAG AGG CAG AAG GCA ATG CAG GTG GAC 1440  
Pro Pro Lys Asp Phe Ala His Lys Arg Gln Lys Ala MET Gln Val Asp  
465 470 475  
40 GCC ACA GTC TTC ACT TGC ATC AGC ATG CTG GCT CGA GCA ATG GGG CCA 1488  
Ala Thr Val Phe Thr Cys Ile Ser MET Leu Ala Arg Ala MET Gly Pro  
480 485 490 495  
45 GGC ATC CAG CAG GAT ATC AAG GAG CTG CTG GAG CCC ATG CTG GCA GTG 1536  
Gly Ile Gln Gln Asp Ile Lys Glu Leu Leu Glu Pro MET Leu Ala Val  
500 505 510  
50 GGA CTA AGC CCT GCC CTC ACT GCA GTG CTC TAC GAC CTG AGC CGT CAG 1584  
Gly Leu Ser Pro Ala Leu Thr Ala Val Leu Tyr Asp Leu Ser Arg Gln  
515 520 525  
55 ATT CCA CAG CTA AAG AAG GAC ATT CAA GAT GGG CTA CTG AAA ATG CTG 1632  
Ile Pro Gln Leu Lys Lys Asp Ile Gln Asp Gly Leu Leu Lys MET Leu  
530 535 540

	TCC CTG GTC CTT ATG CAC AAA CCC CTT CGC CAC CCA GGC ATG CCC AAG	1680
	Ser Leu Val Leu MET His Lys Pro Leu Arg His Pro Gly MET Pro Lys	
	545 550 555	
5	GCC CTG GCC CAT CAG CTG GCC TCT CCT GGC CTC ACG ACC CTC CCT GAG	1728
	Gly Leu Ala His Gln Leu Ala Ser Pro Gly Leu Thr Thr Leu Pro Glu	
	560 565 570 575	
10	GCC AGC GAT GTG GGC AGC ATC ACT CTT GCC CTC CGA ACG CTT GGC AGC	1776
	Ala Ser Asp Val Gly Ser Ile Thr Leu Ala Leu Arg Thr Leu Gly Ser	
	580 585 590	
15	TTT GAA TTT GAA GGC CAC TCT CTG ACC CAA TTT GTT CGC CAC TGT GCG	1824
	Phe Glu Phe Glu Gly His Ser Leu Thr Gln Phe Val Arg His Cys Ala	
	595 600 605	
20	GAT CAT TTC CTG AAC AGT GAG CAC AAG GAG ATC CGC ATG GAG GCT GCC	1872
	Asp His Phe Leu Asn Ser Glu His Lys Glu Ile Arg MET Glu Ala Ala	
	610 615 620	
	CGC ACC TGC TCC CGC CTG CTC ACA CCC TCC ATC CAC CTC ATC AGT GGC	1920
	Arg Thr Cys Ser Arg Leu Leu Thr Pro Ser Ile His Leu Ile Ser Gly	
	625 630 635	
25	CAT GCT CAT GTG GTT AGC CAG ACC GCA GTG CAA GTG GTG GCA GAT GTG	1968
	His Ala His Val Val Ser Gln Thr Ala Val Gln Val Val Ala Asp Val	
	640 645 650 655	
30	CTT AGC AAA CTG CTC GTA GTT GGG ATA ACA GAT CCT GAC CCT GAC ATT	2016
	Leu Ser Lys Leu Leu Val Val Gly Ile Thr Asp Pro Asp Pro Asp Ile	
	660 665 670	
35	CGC TAC TGT GTC TTG GCG TCC CTG GAC GAG CGC TTT GAT GCA CAC CTG	2064
	Arg Tyr Cys Val Leu Ala Ser Leu Asp Glu Arg Phe Asp Ala His Leu	
	675 680 685	
40	GCC CAG GCG GAG AAC TTG CAG GCC TTG TTT GTG GCT CTG AAT GAC CAG	2112
	Ala Gln Ala Glu Asn Leu Gln Ala Leu Phe Val Ala Leu Asn Asp Gln	
	690 695 700	
	GTG TTT GAG ATC CGG GAG CTG GCC ATC TGC ACT GTG GGC CGA CTC AGT	2160
	Val Phe Glu Ile Arg Glu Leu Ala Ile Cys Thr Val Gly Arg Leu Ser	
	705 710 715	
45	AGC ATG AAC CCT GCC TTT GTC ATG CCT TTC CTG CGC AAG ATG CTC ATC	2208
	Ser MET Asn Pro Ala Phe Val MET Pro Phe Leu Arg Lys MET Leu Ile	
	720 725 730 735	
50	CAG ATT TTG ACA GAG TTG GAG CAC AGT GGG ATT GGA AGA ATC AAA GAG	2256
	Gln Ile Leu Thr Glu Leu Glu His Ser Gly Ile Gly Arg Ile Lys Glu	
	740 745 750	

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CAG AGT GCC CGC ATG CTG GGG CAC CTG GTC TCC AAT GCC CCC CGA CTC 2304  
Gln Ser Ala Arg MET Leu Gly His Leu Val Ser Asn Ala Pro Arg Leu  
755 760 765

5 ATC CGC CCC TAC ATG GAG CCT ATT CTG AAG GCA TTA ATT TTG AAA CTG 2352  
Ile Arg Pro Tyr MET Glu Pro Ile Leu Lys Ala Leu Ile Leu Lys Leu  
770 775 780

AAA GAT CCA GAC CCT GAT CCA AAC CCA GGT GTG ATC AAT AAT GTC CTG 2400  
10 Lys Asp Pro Asp Pro Asp Pro Asn Pro Gly Val Ile Asn Asn Val Leu  
785 790 795

GCA ACA ATA GGA GAA TTG GCA CAG GTT AGT GGC CTG GAA ATG AGG AAA 2448  
Ala Thr Ile Gly Glu Leu Ala Gln Val Ser Gly Leu Glu MET Arg Lys  
15 800 805 810 815

TGG GTT GAT GAA CTT TTT ATT ATC ATC ATG GAC ATG CTC CAG GAT TCC 2496  
Trp Val Asp Glu Leu Phe Ile Ile MET Asp MET Leu Gln Asp Ser  
820 825 830

20 TCT TTG TTG GCC AAA AGG CAG GTG GCT CTG TGG ACC CTG GGA CAG TTG 2544  
Ser Leu Leu Ala Lys Arg Gln Val Ala Leu Trp Thr Leu Gly Gln Leu  
835 840 845

25 GTG GCC AGC ACT GGC TAT GTA GTA GAG CCC TAC AGG AAG TAC CCT ACT 2592  
Val Ala Ser Thr Gly Tyr Val Val Glu Pro Tyr Arg Lys Tyr Pro Thr  
850 855 860

30 TTG CTT GAG GTG CTA CTG AAT TTT CTG AAG ACT GAG CAG AAC CAG GGT 2640  
Leu Leu Glu Val Leu Leu Asn Phe Leu Lys Thr Glu Gln Asn Gln Gly  
865 870 875

35 ACA CGC AGA GAG GCC ATC CGT GTG TTA GGG CTT TTA GGG GCT TTG GAT 2688  
Thr Arg Arg Glu Ala Ile Arg Val Leu Gly Leu Leu Gly Ala Leu Asp  
880 885 890 895

40 CCT TAC AAG CAC AAA GTG AAC ATT GGC ATG ATA GAC CAG TCC CGG GAT 2736  
Pro Tyr Lys His Lys Val Asn Ile Gly MET Ile Asp Gln Ser Arg Asp  
900 905 910

45 GCC TCT GCT GTC AGC CTG TCA GAA TCC AAG TCA AGT CAG GAT TCC TCT 2784  
Ala Ser Ala Val Ser Leu Ser Glu Ser Lys Ser Ser Gln Asp Ser Ser  
915 920 925

50 GAC TAT AGC ACT AGT GAA ATG CTG GTC AAC ATG GGA AAC TTG CCT CTG 2832  
Asp Tyr Ser Thr Ser Glu MET Leu Val Asn MET Gly Asn Leu Pro Leu  
930 935 940

GAT GAG TTC TAC CCA GCT GTG TCC ATG GTG GCC CTG ATG CGG ATC TTC 2880  
Asp Glu Phe Tyr Pro Ala Val Ser MET Val Ala Leu MET Arg Ile Phe  
945 950 955

CGA GAC CAG TCA CTC TCT CAT CAT CAC ACC ATG GTT GTC CAG GCC ATC 2928  
Arg Asp Gln Ser Leu Ser His His His Thr MET Val Val Gln Ala Ile  
960 965 970 975

5 ACC TTC ATC TTC AAG TCC CTG GGA CTC AAA TGT GTG CAG TTC CTG CCC 2976  
Thr Phe Ile Phe Lys Ser Leu Gly Leu Lys Cys Val Gln Phe Leu Pro  
980 985 990

10 CAG GTC ATG CCC ACG TTC CTT AAT GTC ATT CGA GTC TGT GAT GGG GCC 3024  
Gln Val MET Pro Thr Phe Leu Asn Val Ile Arg Val Cys Asp Gly Ala  
995 1000 1005

15 ATC CGG GAA TTT TTG TTC CAG CAG CTG GGA ATG TTG GTG TCC TTT GTG 3072  
Ile Arg Glu Phe Leu Phe Gln Gln Leu Gly MET Leu Val Ser Phe Val  
1010 1015 1020

20 AAG AGC CAC ATC AGA CCT TAT ATG GAT GAA ATA GTC ACC CTC ATG AGA 3120  
Lys Ser His Ile Arg Pro Tyr MET Asp Glu Ile Val Thr Leu MET Arg  
1025 1030 1035

25 GAA TTC TGG GTC ATG AAC ACC TCA ATT CAG AGC ACG ATC ATT CTT CTC 3168  
Glu Phe Trp Val MET Asn Thr Ser Ile Gln Ser Thr Ile Ile Leu Leu  
1040 1045 1050 1055

30 ATT GAG CAA ATT GTG GTA GCT CTT GGG GGT GAA TTT AAG CTC TAC CTG 3216  
Ile Glu Gln Ile Val Val Ala Leu Gly Gly Glu Phe Lys Leu Tyr Leu  
1060 1065 1070

35 CCC CAG CTG ATC CCA CAC ATG CTG CGT GTC TTC ATG CAT GAC AAC AGC 3264  
Pro Gln Leu Ile Pro His MET Leu Arg Val Phe MET His Asp Asn Ser  
1075 1080 1085

40 CCA GGC CGC ATT GTC TCT ATC AAG TTA CTG GCT GCA ATC CAG CTG TTT 3312  
Pro Gly Arg Ile Val Ser Ile Lys Leu Leu Ala Ala Ile Gln Leu Phe  
1090 1095 1100

45 GGC GCC AAC CTG GAT GAC TAC CTG CAT TTA CTG CTG CCT CCT ATT GTT 3360  
Gly Ala Asn Leu Asp Asp Tyr Leu His Leu Leu Leu Pro Pro Ile Val  
1105 1110 1115

50 AAG TTG TTT GAT GCC CCT GAA GCT CCA CTG CCA TCT CGA AAG GCA GCG 3408  
Lys Leu Phe Asp Ala Pro Glu Ala Pro Leu Pro Ser Arg Lys Ala Ala  
1120 1125 1130 1135

55 CTA GAG ACT GTG GAC CGC CTG ACG GAG TCC CTG GAT TTC ACT GAC TAT 3456  
Leu Glu Thr Val Asp Arg Leu Thr Glu Ser Leu Asp Phe Thr Asp Tyr  
1140 1145 1150

50 GCC TCC CGG ATC ATT CAC CCT ATT GTT CGA ACA CTG GAC CAG AGC CCA 3504  
Ala Ser Arg Ile Ile His Pro Ile Val Arg Thr Leu Asp Gln Ser Pro  
1155 1160 1165

GAA CTG CGC TCC ACA GCC ATG GAC ACG CTG TCT TCA CTT GTT TTT CAG 3552  
Glu Leu Arg Ser Thr Ala MET Asp Thr Leu Ser Ser Leu Val Phe Gln  
1170 1175 1180

5 CTG GGG AAG AAG TAC CAA ATT TTC ATT CCA ATG GTG AAT AAA GTT CTG 3600  
Leu Gly Lys Lys Tyr Gln Ile Phe Ile Pro MET Val Asn Lys Val Leu  
1185 1190 1195

10 GTG CGA CAC CGA ATC AAT CAT CAG CGC TAT GAT GTG CTC ATC TGC AGA 3648  
Val Arg His Arg Ile Asn His Gln Arg Tyr Asp Val Leu Ile Cys Arg  
1200 1205 1210 1215

15 ATT GTC AAG GGA TAC ACA CTT GCT GAT GAA GAG GAG GAT CCT TTG ATT 3696  
Ile Val Lys Gly Tyr Thr Leu Ala Asp Glu Glu Asp Pro Leu Ile  
1220 1225 1230

20 TAC CAG CAT CGG ATG CTT AGG AGT GGC CAA GGG GAT GCA TTG GCT AGT 3744  
Tyr Gln His Arg MET Leu Arg Ser Gly Gln Gly Asp Ala Leu Ala Ser  
1235 1240 1245

25 GGA CCA GTG GAA ACA GGA CCC ATG AAG AAA CTG CAC GTC AGC ACC ATC 3792  
Gly Pro Val Glu Thr Gly Pro MET Lys Lys Leu His Val Ser Thr Ile  
1250 1255 1260

30 AAC CTC CAA AAG GCC TGG GGC GCT GCC AGG AGG GTC TCC AAA GAT GAC 3840  
Asn Leu Gln Lys Ala Trp Gly Ala Ala Arg Arg Val Ser Lys Asp Asp  
1265 1270 1275

35 TGG CTG GAA TGG CTG AGA CGG CTG AGC CTG GAG CTG CTG AAG GAC TCA 3888  
Trp Leu Glu Trp Leu Arg Arg Leu Ser Leu Glu Leu Leu Lys Asp Ser  
1280 1285 1290 1295

40 TCA TCG CCC TCC CTG CGC TCC TGC TGG GCC CTG GCA CAG GCC TAC AAC 3936  
Ser Ser Pro Ser Leu Arg Ser Cys Trp Ala Leu Ala Gln Ala Tyr Asn  
1300 1305 1310

45 CCG ATG GCC AGG GAT CTC TTC AAT GCT GCA TTT GTG TCC TGC TGG TCT 3984  
Pro MET Ala Arg Asp Leu Phe Asn Ala Ala Phe Val Ser Cys Trp Ser  
1315 1320 1325

50 GAA CTG AAT GAA GAT CAA CAG GAT GAG CTC ATC AGA AGC ATC GAG TTG 4032  
Glu Leu Asn Glu Asp Gln Gln Asp Glu Leu Ile Arg Ser Ile Glu Leu  
1330 1335 1340

45 GCC CTC ACC TCA CAA GAC ATC GCT GAA GTC ACA CAG ACC CTC TTA AAC 4080  
Ala Leu Thr Ser Gln Asp Ile Ala Glu Val Thr Gln Thr Leu Leu Asn  
1345 1350 1355

50 TTG GCT GAA TTC ATG GAA CAC AGT GAC AAG GGC CCC CTG CCA CTG AGA 4128  
Leu Ala Glu Phe MET Glu His Ser Asp Lys Gly Pro Leu Pro Leu Arg  
1360 1365 1370 1375

	GAT GAC AAT GGC ATT GTT CTG CTG GGT GAG AGA GCT GCC AAG TGC CGA	4176
	Asp Asp Asn Gly Ile Val Leu Leu Gly Glu Arg Ala Ala Lys Cys Arg	
	1380 1385 1390	
5	GCA TAT GCC AAA GCA CTA CAC TAC AAA GAA CTG GAG TTC CAG AAA GGC	4224
	Ala Tyr Ala Lys Ala Leu His Tyr Lys Glu Leu Glu Phe Gln Lys Gly	
	1395 1400 1405	
10	CCC ACC CCT GCC ATT CTA GAA TCT CTC ATC AGC ATT AAT AAT AAG CTA	4272
	Pro Thr Pro Ala Ile Leu Glu Ser Leu Ile Ser Ile Asn Asn Lys Leu	
	1410 1415 1420	
15	CAG CAG CCG GAG GCA GCG GCC GGA GTG TTA GAA TAT GCC ATG AAA CAC	4320
	Gln Gln Pro Glu Ala Ala Gly Val Leu Glu Tyr Ala MET Lys His	
	1425 1430 1435	
20	TTT GGA GAG CTG GAG ATC CAG GCT ACC TGG TAT GAG AAA CTG CAC GAG	4368
	Phe Gly Glu Leu Glu Ile Gln Ala Thr Trp Tyr Glu Lys Leu His Glu	
	1440 1445 1450 1455	
	TGG GAG GAT GCC CTT GTG GCC TAT GAC AAG AAA ATG GAC ACC AAC AAG	4416
	Trp Glu Asp Ala Leu Val Ala Tyr Asp Lys Lys MET Asp Thr Asn Lys	
	1460 1465 1470	
25	GAC GAC CCA GAG CTG ATG CTG GGC CGC ATG CGC TGC CTC GAG GCC TTG	4464
	Asp Asp Pro Glu Leu MET Leu Gly Arg MET Arg Cys Leu Glu Ala Leu	
	1475 1480 1485	
30	GGG GAA TGG GGT CAA CTC CAC CAG CAG TGC TGT GAA AAG TGG ACC CTG	4512
	Gly Glu Trp Gly Gln Leu His Gln Gln Cys Cys Glu Lys Trp Thr Leu	
	1490 1495 1500	
35	GTT AAT GAT GAG ACC CAA GCC AAG ATG GCC CGG ATG GCT GCT GCA GCT	4560
	Val Asn Asp Glu Thr Gln Ala Lys MET Ala Arg MET Ala Ala Ala Ala	
	1505 1510 1515	
40	GCA TGG GGT TTA GGT CAG TGG GAC AGC ATG GAA GAA TAC ACC TGT ATG	4608
	Ala Trp Gly Leu Gly Gln Trp Asp Ser MET Glu Glu Tyr Thr Cys MET	
	1520 1525 1530 1535	
	ATC CCT CGG GAC ACC CAT GAT GGG GCA TTT TAT AGA GCT GTG CTG GCA	4656
	Ile Pro Arg Asp Thr His Asp Gly Ala Phe Tyr Arg Ala Val Leu Ala	
	1540 1545 1550	
45	CTG CAT CAG GAC CTC TTC TCC TTG GCA CAA CAG TGC ATT GAC AAG GCC	4704
	Leu His Gln Asp Leu Phe Ser Leu Ala Gln Gln Cys Ile Asp Lys Ala	
	1555 1560 1565	
50	AGG GAC CTG CTG GAT GCT GAA TTA ACT GCA ATG GCA GGA GAG AGT TAC	4752
	Arg Asp Leu Leu Asp Ala Glu Leu Thr Ala MET Ala Gly Glu Ser Tyr	
	1570 1575 1580	

	AGT CGG GCA TAT GGG GCC ATG GTT TCT TGC CAC ATG CTG TCC GAG CTG	4800
	Ser Arg Ala Tyr Gly Ala MET Val Ser Cys His MET Leu Ser Glu Leu	
	1585 1590 1595	
5	GAG GAG GTT ATC CAG TAC AAA CTT GTC CCC GAG CGA CGA GAG ATC ATC	4848
	Glu Glu Val Ile Gln Tyr Lys Leu Val Pro Glu Arg Arg Glu Ile Ile	
	1600 1605 1610 1615	
10	CGC CAG ATC TGG TGG GAG AGA CTG CAG GGC TGC CAG CGT ATC GTA GAG	4896
	Arg Gln Ile Trp Trp Glu Arg Leu Gln Gly Cys Gln Arg Ile Val Glu	
	1620 1625 1630	
15	GAC TGG CAG AAA ATC CTT ATG GTG CGG TCC CTT GTG GTC AGC CCT CAT	4944
	Asp Trp Gln Lys Ile Leu MET Val Arg Ser Leu Val Val Ser Pro His	
	1635 1640 1645	
20	GAA GAC ATG AGA ACC TGG CTC AAG TAT GCA AGC CTG TGC GGC AAG AGT	4992
	Glu Asp MET Arg Thr Trp Leu Lys Tyr Ala Ser Leu Cys Gly Lys Ser	
	1650 1655 1660	
	GGC AGG CTG GCT CTT GCT CAT AAA ACT TTA GTG TTG CTC CTG GGA GTT	5040
	Gly Arg Leu Ala Leu Ala His Lys Thr Leu Val Leu Leu Leu Gly Val	
	1665 1670 1675	
25	GAT CCG TCT CGG CAA CTT GAC CAT CCT CTG CCA ACA GTT CAC CCT CAG	5088
	Asp Pro Ser Arg Gln Leu Asp His Pro Leu Pro Thr Val His Pro Gln	
	1680 1685 1690 1695	
30	GTG ACC TAT GCC TAC ATG AAA AAC ATG TGG AAG AGT GCC CGC AAG ATC	5136
	Val Thr Tyr Ala Tyr MET Lys Asn MET Trp Lys Ser Ala Arg Lys Ile	
	1700 1705 1710	
35	GAT GCC TTC CAG CAC ATG CAG CAT TTT GTC CAG ACC ATG CAG CAA CAG	5184
	Asp Ala Phe Gln His MET Gln His Phe Val Gln Thr MET Gln Gln Gln	
	1715 1720 1725	
40	GCC CAG CAT GCC ATC GCT ACT GAG GAC CAG CAG CAT AAG CAG GAA CTG	5232
	Ala Gln His Ala Ile Ala Thr Glu Asp Gln Gln His Lys Gln Glu Leu	
	1730 1735 1740	
	CAC AAG CTC ATG GCC CGA TGC TTC CTG AAA CTT GGA GAG TGG CAG CTG	5280
	His Lys Leu MET Ala Arg Cys Phe Leu Lys Leu Gly Glu Trp Gln Leu	
	1745 1750 1755	
45	AAT CTA CAG GGC ATC AAT GAG AGC ACA ATC CCC AAA GTG CTG CAG TAC	5328
	Asn Leu Gln Gly Ile Asn Glu Ser Thr Ile Pro Lys Val Leu Gln Tyr	
	1760 1765 1770 1775	
50	TAC AGC GCC GCC ACA GAG CAC GAC CGC AGC TGG TAC AAG GCC TGG CAT	5376
	Tyr Ser Ala Ala Thr Glu His Asp Arg Ser Trp Tyr Lys Ala Trp His	
	1780 1785 1790	

	GCG TGG GCA GTG ATG AAC TTC GAA GCT GTG CTA CAC TAC AAA CAT CAG	' 5424
	Ala Trp Ala Val MET Asn Phe Glu Ala Val Leu His Tyr Lys His Gln	
	1795 1800 1805	
5	AAC CAA GCC CGC GAT GAG AAG AAA CTG CGT CAT GCC AGC GGG GCC	5472
	Asn Gln Ala Arg Asp Glu Lys Lys Leu Arg His Ala Ser Gly Ala	
	1810 1815 1820	
10	AAC ATC ACC AAC GCC ACC ACT GCC GCC ACC ACG GCC GCC ACT GCC ACC	5520
	Asn Ile Thr Asn Ala Thr Thr Ala Ala Thr Thr Ala Ala Thr Ala Thr	
	1825 1830 1835	
15	ACC ACT GCC AGC ACC GAG GGC AGC AAC AGT GAG AGC GAG GCC GAG AGC	5568
	Thr Thr Ala Ser Thr Glu Gly Ser Asn Ser Glu Ser Glu Ala Glu Ser	
	1840 1845 1850 1855	
	ACC GAG AAC AGC CCC ACC CCA TCG CCG CTG CAG AAG AAG GTC ACT GAG	5616
	Thr Glu Asn Ser Pro Thr Pro Ser Pro Leu Gln Lys Lys Val Thr Glu	
	1860 1865 1870	
20	GAT CTG TCC AAA ACC CTC CTG ATG TAC ACG GTG CCT GCC GTC CAG GGC	5664
	Asp Leu Ser Lys Thr Leu Leu MET Tyr Thr Val Pro Ala Val Gln Gly	
	1875 1880 1885	
25	TTC TTC CGT TCC ATC TCC TTG TCA CGA GGC AAC AAC CTC CAG GAT ACA	5712
	Phe Phe Arg Ser Ile Ser Leu Ser Arg Gly Asn Asn Leu Gln Asp Thr	
	1890 1895 1900	
30	CTC AGA GTT CTC ACC TTA TGG TTT GAT TAT GGT CAC TGG CCA GAT GTC	5760
	Leu Arg Val Leu Thr Leu Trp Phe Asp Tyr Gly His Trp Pro Asp Val	
	1905 1910 1915	
35	AAT GAG GCC TTA GTG GAG GGG GTG AAA GCC ATC CAG ATT GAT ACC TGG	5808
	Asn Glu Ala Leu Val Glu Gly Val Lys Ala Ile Gln Ile Asp Thr Trp	
	1920 1925 1930 1935	
	CTA CAG GTT ATA CCT CAG CTC ATT GCA AGA ATT GAT ACG CCC AGA CCC	5856
	Leu Gln Val Ile Pro Gln Leu Ile Ala Arg Ile Asp Thr Pro Arg Pro	
	1940 1945 1950	
40	TTG GTG GGA CGT CTC ATT CAC CAG CTT CTC ACA GAC ATT GGT CGG TAC	5904
	Leu Val Gly Arg Leu Ile His Gln Leu Leu Thr Asp Ile Gly Arg Tyr	
	1955 1960 1965	
45	CAC CCC CAG GCC CTC ATC TAC CCA CTG ACA GTG GCT TCT AAG TCT ACC	5952
	His Pro Gln Ala Leu Ile Tyr Pro Leu Thr Val Ala Ser Lys Ser Thr	
	1970 1975 1980	
50	ACG ACA GCC CGG CAC AAT GCA GCC AAC AAG ATT CTG AAG AAC ATG TGT	6000
	Thr Thr Ala Arg His Asn Ala Ala Asn Lys Ile Leu Lys Asn MET Cys	
	1985 1990 1995	

	GAG CAC AGC AAC ACC CTG GTC CAG CAG GCC ATG ATG GTG AGC GAG GAG	6048
	Glu His Ser Asn Thr Leu Val Gln Gln Ala MET MET Val Ser Glu Glu	
	2000 2005 2010 2015	
5	CTG ATC CGA GTG GCC ATC CTC TGG CAT GAG ATG TGG CAT GAA GGC CTG	6096
	Leu Ile Arg Val Ala Ile Leu Trp His Glu MET Trp His Glu Gly Leu	
	2020 2025 2030	
10	GAA GAG GCA TCT CGT TTG TAC TTT GGG GAA AGG AAC GTG AAA GGC ATG	6144
	Glu Glu Ala Ser Arg Leu Tyr Phe Gly Glu Arg Asn Val Lys Gly MET	
	2035 2040 2045	
15	TTT GAG GTG CTG GAG CCC TTG CAT GCT ATG ATG GAA CGG GGC CCC CAG	6192
	Phe Glu Val Leu Glu Pro Leu His Ala MET MET Glu Arg Gly Pro Gln	
	2050 2055 2060	
	ACT CTG AAG GAA ACA TCC TTT AAT CAG GCC TAT GGT CGA GAT TTA ATG	6240
	Thr Leu Lys Glu Thr Ser Phe Asn Gln Ala Tyr Gly Arg Asp Leu MET	
	2065 2070 2075	
20	GAG GCC CAA GAG TGG TGC AGG AAG TAC ATG AAA TCA GGG AAT GTC AAG	6288
	Glu Ala Gln Glu Trp Cys Arg Lys Tyr MET Lys Ser Gly Asn Val Lys	
	2080 2085 2090 2095	
25	GAC CTC ACC CAA GCC TGG GAC CTC TAT TAT CAT GTG TTC CGA CGA ATC	6336
	Asp Leu Thr Gln Ala Trp Asp Leu Tyr Tyr His Val Phe Arg Arg Ile	
	2100 2105 2110	
30	TCA AAG CAG CTG CCT CAG CTC ACA TCC TTA GAG CTG CAA TAT GTT TCC	6384
	Ser Lys Gln Leu Pro Gln Leu Thr Ser Leu Glu Leu Gln Tyr Val Ser	
	2115 2120 2125	
35	CCA AAA CTT CTG ATG TGC CGG GAC CTT GAA TTG GCT GTG CCA GGA ACA	6432
	Pro Lys Leu Leu MET Cys Arg Asp Leu Glu Leu Ala Val Pro Gly Thr	
	2130 2135 2140	
	TAT GAC CCC AAC CAG CCA ATC ATT CGC ATT CAG TCC ATA GCA CCG TCT	6480
	Tyr Asp Pro Asn Gln Pro Ile Ile Arg Ile Gln Ser Ile Ala Pro Ser	
	2145 2150 2155	
40	TTG CAA GTC ATC ACA TCC AAG CAG AGG CCC CGG AAA TTG ACA CTT ATG	6528
	Leu Gln Val Ile Thr Ser Lys Gln Arg Pro Arg Lys Leu Thr Leu MET	
	2160 2165 2170 2175	
45	GGC AGC AAC GGA CAT GAG TTT GTT TTC CTT CTA AAA GGC CAT GAA GAT	6576
	Gly Ser Asn Gly His Glu Phe Val Phe Leu Leu Lys Gly His Glu Asp	
	2180 2185 2190	
50	CTG CGC CAG GAT GAG CGT GTG ATG CAG CTC TTC GGC CTG GTT AAC ACC	6624
	Leu Arg Gln Asp Glu Arg Val MET Gln Leu Phe Gly Leu Val Asn Thr	
	2195 2200 2205	

- 40 -

CTT CTG GCC AAT GAC CCA ACA TCT CTT CGG AAA AAC CTC AGC ATC CAG 6672  
Leu Leu Ala Asn Asp Pro Thr Ser Leu Arg Lys Asn Leu Ser Ile Gln  
2210 2215 2220

5 AGA TAC GCT GTC ATC CCT TTA TCG ACC AAC TCG GGC CTC ATT GGC TGG 6720  
Arg Tyr Ala Val Ile Pro Leu Ser Thr Asn Ser Gly Leu Ile Gly Trp  
2225 2230 2235

10 GTT CCC CAC TGT GAC ACA CTG CAC GCC CTC ATC CGG GAC TAC AGG GAG 6768  
Val Pro His Cys Asp Thr Leu His Ala Leu Ile Arg Asp Tyr Arg Glu  
2240 2245 2250 2255

15 AAG AAG AAG ATC CTT CTC AAC ATC GAG CAT CGC ATC ATG TTG CGG ATG 6816  
Lys Lys Lys Ile Leu Leu Asn Ile Glu His Arg Ile MET Leu Arg MET  
2260 2265 2270

GCT CCG GAC TAT GAC CAC TTG ACT CTG ATG CAG AAG GTG GAG GTG TTT 6864  
Ala Pro Asp Tyr Asp His Leu Thr Leu MET Gln Lys Val Glu Val Phe  
2275 2280 2285

20 GAG CAT GCC GTC AAT AAT ACA GCT GGG GAC GAC CTG GCC AAG CTG CTG 6912  
Glu His Ala Val Asn Asn Thr Ala Gly Asp Asp Leu Ala Lys Leu Leu  
2290 2295 2300

25 TGG CTG AAA AGC CCC AGC TCC GAG GTG TGG TTT GAC CGA AGA ACC AAT 6960  
Trp Leu Lys Ser Pro Ser Ser Glu Val Trp Phe Asp Arg Arg Thr Asn  
2305 2310 2315

30 TAT ACC CGT TCT TTA GCG GTC ATG TCA ATG GTT GGG TAT ATT TTA GGC 7008  
Tyr Thr Arg Ser Leu Ala Val MET Ser MET Val Gly Tyr Ile Leu Gly  
2320 2325 2330 2335

35 CTG GGA GAT AGA CAC CCA TCC AAC CTG ATG CTG GAC CGT CTG AGT GGG 7056  
Leu Gly Asp Arg His Pro Ser Asn Leu MET Leu Asp Arg Leu Ser Gly  
2340 2345 2350

40 AAG ATC CTG CAC ATT GAC TTT GGG GAC TGC TTT GAG GTT GCT ATG ACC 7104  
Lys Ile Leu His Ile Asp Phe Gly Asp Cys Phe Glu Val Ala MET Thr  
2355 2360 2365

45 CGA GAG AAG TTT CCA GAG AAG ATT CCA TTT AGA CTA ACA AGA ATG TTG 7152  
Arg Glu Lys Phe Pro Glu Lys Ile Pro Phe Arg Leu Thr Arg MET Leu  
2370 2375 2380

50 ACC AAT GCT ATG GAG GTT ACA GGC CTG GAT GGC AAC TAC AGA ATC ACA 7200  
Thr Asn Ala MET Glu Val Thr Gly Leu Asp Gly Asn Tyr Arg Ile Thr  
2385 2390 2395

TGC CAC ACA GTG ATG GAG GTG CTG CGA GAG CAC AAG GAC AGT GTC ATG 7248  
Cys His Thr Val MET Glu Val Leu Arg Glu His Lys Asp Ser Val MET  
2400 2405 2410 2415

GCC GTG CTG GAA GCC TTT GTC TAT GAC CCC TTG CTG AAC TGG AGG CTG 7296  
Ala Val Leu Glu Ala Phe Val Tyr Asp Pro Leu Leu Asn Trp Arg Leu  
2420 2425 2430

5 ATG GAC ACA AAT ACC AAA GGC AAC AAG CGA TCC CGA ACG AGG ACG GAT 7344  
MET Asp Thr Asn Thr Lys Gly Asn Lys Arg Ser Arg Thr Arg Thr Asp  
2435 2440 2445

TCC TAC TCT GCT GGC CAG TCA GTC GAA ATT TTG GAC GGT GTG GAA CTT 7392  
10 Ser Tyr Ser Ala Gly Gln Ser Val Glu Ile Leu Asp Gly Val Glu Leu  
2450 2455 2460

GGA GAG CCA GCC CAT AAG AAA ACG GGG ACC ACA GTG CCA GAA TCT ATT 7440  
Gly Glu Pro Ala His Lys Lys Thr Gly Thr Thr Val Pro Glu Ser Ile  
15 2465 2470 2475

CAT TCT TTC ATT GGA GAC GGT TTG GTG AAA CCA GAG GCC CTA AAT AAG 7488  
His Ser Phe Ile Gly Asp Gly Leu Val Lys Pro Glu Ala Leu Asn Lys  
2480 2485 2490 2495

20 AAA GCT ATC CAG ATT ATT AAC AGG GTT CGA GAT AAG CTC ACT GGT CGG . 7536  
Lys Ala Ile Gln Ile Ile Asn Arg Val Arg Asp Lys Leu Thr Gly Arg  
2500 2505 2510

25 GAC TTC TCT CAT GAT GAC ACT TTG GAT GTT CCA ACG CAA GTT GAG CTG 7584  
Asp Phe Ser His Asp Asp Thr Leu Asp Val Pro Thr Gln Val Glu Leu  
2515 2520 2525

CTC ATC AAA CAA GCG ACA TCC CAT GAA AAC CTC TGC CAG TGC TAT ATT 7632  
30 Leu Ile Lys Gln Ala Thr Ser His Glu Asn Leu Cys Gln Cys Tyr Ile  
2530 2535 2540

GGC TGG TAC CCT TTC TGG TAA 7653  
Gly Trp Tyr Pro Phe Trp  
35 2545

(3) INFORMATION FOR SEQ. ID NO: 2:  
(i) SEQUENCE CHARACTERISTICS:  
(A) LENGTH: 3423  
40 (B) TYPE: nucleic acid  
(C) STRANDEDNESS: double-stranded  
(D) TOPOLOGY: linear

45 (ii) MOLECULE TYPE: sense orientation of double-stranded  
cDNA to mRNA  
(A) DESCRIPTION: Sequence No. 2 illustrates a  
GST-SEP45 fusion protein beginning  
at the first amino acid of the GST-SEP45  
protein.

50 (iii) HYPOTHETICAL: no  
(iv) ANTISENSE: no  
(vi) ORIGINAL SOURCE:  
(A) ORGANISM: Molt 4 human T-cell leukemia cells  
55 (B) STRAIN: ATCC Strain CRL 1582

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(xi) SEQUENCE DESCRIPTION: SEQ. ID NO: 2

	ATG TCC CCT ATA CTA GGT TAT TGG AAA ATT AAG GGC CTT GTG CAA CCC	48
5	MET Ser Pro Ile Leu Gly Tyr Trp Lys Ile Lys Gly Leu Val Gln Pro	
	1 5 10 15	
	ACT CGA CTT CTT TTG GAA TAT CTT GAA GAA AAA TAT GAA GAG CAT TTG	96
	Thr Arg Leu Leu Leu Glu Tyr Leu Glu Glu Lys Tyr Glu Glu His Leu	
10	20 25 30	
	TAT GAG CGC GAT GAA GGT GAT AAA TGG CGA AAC AAA AAG TTT GAA TTG	144
	Tyr Glu Arg Asp Glu Gly Asp Lys Trp Arg Asn Lys Lys Phe Glu Leu	
	35 40 45	
15	GGT TTG GAG TTT CCC AAT CTT CCT TAT TAT ATT GAT GGT GAT GTT AAA	192
	Gly Leu Glu Phe Pro Asn Leu Pro Tyr Tyr Ile Asp Gly Asp Val Lys	
	50 55 60	
20	TTA ACA CAG TCT ATG GCC ATC ATA CGT TAT ATA GCT GAC AAG CAC AAC	240
	Leu Thr Gln Ser MET Ala Ile Ile Arg Tyr Ile Ala Asp Lys His Asn	
	65 70 75 80	
25	ATG TTG GGT TGT CCA AAA GAG CGT GCA GAG ATT TCA ATG CTT GAA	288
	MET Leu Gly Gly Cys Pro Lys Glu Arg Ala Glu Ile Ser MET Leu Glu	
	85 90 95	
30	GGA GCG GTT TTG GAT ATT AGA TAC GGT GTT TCG AGA ATT GCA TAT AGT	336
	Gly Ala Val Leu Asp Ile Arg Tyr Gly Val Ser Arg Ile Ala Tyr Ser	
	100 105 110	
	AAA GAC TTT GAA ACT CTC AAA GTT GAT TTT CTT AGC AAG CTA CCT GAA	384
	Lys Asp Phe Glu Thr Leu Lys Val Asp Phe Leu Ser Lys Leu Pro Glu	
	115 120 125	
35	ATG CTG AAA ATG TTC GAA GAT CGT TTA TGT CAT AAA ACA TAT TTA AAT	432
	MET Leu Lys MET Phe Glu Asp Arg Leu Cys His Lys Thr Tyr Leu Asn	
	130 135 140	
40	GGT GAT CAT GTA ACC CAT CCT GAC TTC ATG TTG TAT GAC GCT CTT GAT	480
	Gly Asp His Val Thr His Pro Asp Phe MET Leu Tyr Asp Ala Leu Asp	
	145 150 155 160	
45	GTT GTT TTA TAC ATG GAC CCA ATG TGC CTG GAT GCG TTC CCA AAA TTA	528
	Val Val Leu Tyr MET Asp Pro MET Cys Leu Asp Ala Phe Pro Lys Leu	
	165 170 175	
	GTT TGT TTT AAA AAA CGT ATT GAA GCT ATC CCA CAA ATT GAT AAG TAC	576
	Val Cys Phe Lys Lys Arg Ile Glu Ala Ile Pro Gln Ile Asp Lys Tyr	
	180 185 190	
50	TTG AAA TCC AGC AAG TAT ATA GCA TGG CCT TTG CAG GGC TGG CAA GCC	624
	Leu Lys Ser Ser Lys Tyr Ile Ala Trp Pro Leu Gln Gly Trp Gln Ala	
	195 200 205	

ACG TTT GGT GGT GGC GAC CAT CCT CCA AAA TCG GAT CTG GTT CCG CGT' 672  
Thr Phe Gly Gly Gly Asp His Pro Pro Lys Ser Asp Leu Val Pro Arg  
210 215 220

5 GGT GGA TCC CCG GGA ATT TCC GGT GGT GGT GGA ATT CTA GAC 720  
Gly Gly Ser Pro Gly Ile Ser Gly Gly Gly Gly Ile Leu Asp  
225 230 235 240

10 GAC TCC ATG AGC TTC AAG TAT GCA AGC CTG TGC GGC AAG AGT GGC AGG 768  
Asp Ser MET Ser Phe Lys Tyr Ala Ser Leu Cys Gly Lys Ser Gly Arg  
245 250 255

15 CTG GCT CTT GCT CAT AAA ACT TTA GTG TTG CTC CTG GGA GTT GAT CCG' 816  
Leu Ala Leu Ala His Lys Thr Leu Val Leu Leu Leu Gly Val Asp Pro  
260 265 270

20 TCT CGG CAA CTT GAC CAT CCT CTG CCA ACA GTT CAC CCT CAG GTG ACC 864  
Ser Arg Gln Leu Asp His Pro Leu Pro Thr Val His Pro Gln Val Thr  
275 280 285

25 TAT GCC TAC ATG AAA AAC ATG TGG AAG AGT GCC CGC AAG ATC GAT GCC 912  
Tyr Ala Tyr MET Lys Asn MET Trp Lys Ser Ala Arg Lys Ile Asp Ala  
290 295 300

30 TTC CAG CAC ATG CAG CAT TTT GTC CAG ACC ATG CAG CAA CAG GCC CAG 960  
Phe Gln His MET Gln His Phe Val Gln Thr MET Gln Gln Ala Gln  
305 310 315 320

35 CAT GCC ATC GCT ACT GAG GAC CAG CAG CAT AAG CAG GAA CTG CAC AAG 1008  
His Ala Ile Ala Thr Glu Asp Gln Gln His Lys Gln Glu Leu His Lys  
325 330 335

40 CTC ATG GCC CGA TGC TTC CTG AAA CTT GGA GAG TGG CAG CTG AAT CTA 1056  
Leu MET Ala Arg Cys Phe Leu Lys Leu Gly Glu Trp Gln Leu Asn Leu  
340 345 350

45 CAG GGC ATC AAT GAG AGC ACA ATC CCC AAA GTG CTG CAG TAC TAC AGC 1104  
Gln Gly Ile Asn Glu Ser Thr Ile Pro Lys Val Leu Gln Tyr Tyr Ser  
355 360 365

50 GCC GCC ACA GAG CAC GAC CGC AGC TGG TAC AAG GCC TGG CAT GCG TGG 1152  
Ala Ala Thr Glu His Asp Arg Ser Trp Tyr Lys Ala Trp His Ala Trp  
370 375 380

55 GCA GTG ATG AAC TTC GAA GCT GTG CTA CAC TAC AAA CAT CAG AAC CAA 1200  
Ala Val MET Asn Phe Glu Ala Val Leu His Tyr Lys His Gln Asn Gln  
385 390 395 400

60 GCC CGC GAT GAG AAG AAG AAA CTG CGT CAT GCC AGC GGG GCC AAC ATC 1248  
Ala Arg Asp Glu Lys Lys Lys Leu Arg His Ala Ser Gly Ala Asn Ile  
405 410 415

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	ACC AAC GCC ACC ACT GCC GCC ACC ACG GCC GCC ACT GCC ACC ACC ACT	1296
	Thr Asn Ala Thr Thr Ala Ala Thr Thr Ala Ala Thr Ala Thr Thr Thr	
	420	425
	430	
5	GCC AGC ACC GAG GGC AGC AAC AGT GAG AGT GAG GCC GAG AGC ACC GAG	1344
	Ala Ser Thr Glu Gly Ser Asn Ser Glu Ser Glu Ala Glu Ser Thr Glu	
	435	440
	445	
10	AAC AGC CCC ACC CCA TCG CCG CTG CAG AAG AAG GTC ACT GAG GAT CTG	1392
	Asn Ser Pro Thr Pro Ser Pro Leu Gln Lys Lys Val Thr Glu Asp Leu	
	450	455
	460	
15	TCC AAA ACC CTC CTG ATG TAC ACG GTG CCT GCC GTC CAG GGC TTC TTC	1440
	Ser Lys Thr Leu Leu MET Tyr Thr Val Pro Ala Val Gln Gly Phe Phe	
	465	470
	475	480
	485	
20	CGT TCC ATC TCC TTG TCA CGA GGC AAC AAC CTC CAG GAT ACA CTC AGA	1488
	Arg Ser Ile Ser Leu Ser Arg Gly Asn Asn Leu Gln Asp Thr Leu Arg	
	485	490
	495	
25	GTT CTC ACC TTA TGG TTT GAT TAT GGT CAC TGG CCA GAT GTC AAT GAG	1536
	Val Leu Thr Leu Trp Phe Asp Tyr Gly His Trp Pro Asp Val Asn Glu	
	500	505
	510	
30	GCC TTA GTG GAG GGG GTG AAA GCC ATC CAG ATT GAT ACC TGG CTA CAG	1584
	Ala Leu Val Glu Gly Val Lys Ala Ile Gln Ile Asp Thr Trp Leu Gln	
	515	520
	525	
35	GTT ATA CCT CAG CTC ATT GCA AGA ATT GAT ACG CCC AGA CCC TTG GTG	1632
	Val Ile Pro Gln Leu Ile Ala Arg Ile Asp Thr Pro Arg Pro Leu Val	
	530	535
	540	
	545	
	550	555
	560	
	565	
40	GGA CGT CTC ATT CAC CAG CTT CTC ACA GAC ATT GGT CGG TAC CAC CCC	1680
	Gly Arg Leu Ile His Gln Leu Leu Thr Asp Ile Gly Arg Tyr His Pro	
	545	550
	555	560
	565	
	570	
	575	
45	CAG GCC CTC ATC TAC CCA CTG ACA GTG GCT TCT AAG TCT ACC ACG ACA	1728
	Gln Ala Leu Ile Tyr Pro Leu Thr Val Ala Ser Lys Ser Thr Thr Thr	
	565	570
	575	
	580	
	585	590
50	GCC CGG CAC AAT GCA GCC AAC AAG ATT CTG AAG AAC ATG TGT GAG CAC	1776
	Ala Arg His Asn Ala Ala Asn Lys Ile Leu Lys Asn MET Cys Glu His	
	580	585
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	600	605
	610	615
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GCA TCT CGT TTG TAC TTT GGG GAA AGG AAC GTG AAA GGC ATG TTT GAG 1920  
Ala Ser Arg Leu Tyr Phe Gly Glu Arg Asn Val Lys Gly MET Phe Glu  
625 630 635 640

5 GTG CTG GAG CCC TTG CAT GCT ATG ATG GAA CGG GGC CCC CAG ACT CTG 1968  
Val Leu Glu Pro Leu His Ala MET MET Glu Arg Gly Pro Gln Thr Leu  
645 650 655

10 AAG GAA ACA TCC TTT AAT CAG GCC TAT GGT CGA GAT TTA ATG GAG GCC 2016  
Lys Glu Thr Ser Phe Asn Gln Ala Tyr Gly Arg Asp Leu MET Glu Ala  
660 665 670

15 CAA GAG TGG TGC AGG AAG TAC ATG AAA TCA GGG AAT GTC AAG GAC CTC 2064  
Gln Glu Trp Cys Arg Lys Tyr MET Lys Ser Gly Asn Val Lys Asp Leu  
675 680 685

20 ACC CAA GCC TGG GAC CTC TAT TAT CAT GTG TTC CGA CGA ATC TCA AAG 2112  
Thr Gln Ala Trp Asp Leu Tyr Tyr His Val Phe Arg Arg Ile Ser Lys  
690 695 700

25 CAG CTG CCT CAG CTC ACA TCC TTA GAG CTG CAA TAT GTT TCC CCA AAA 2160  
Gln Leu Pro Gln Leu Thr Ser Leu Glu Leu Gln Tyr Val Ser Pro Lys  
705 710 715 720

30 CTT CTG ATG TGC CGG GAC CTT GAA TTG GCT GTG CCA GGA ACA TAT GAC 2208  
Leu Leu MET Cys Arg Asp Leu Glu Leu Ala Val Pro Gly Thr Tyr Asp  
725 730 735

35 CCC AAC CAG CCA ATC ATT CGC ATT CAG TCC ATA GCA CCG TCT TTG CAA 2256  
Pro Asn Gln Pro Ile Ile Arg Ile Gln Ser Ile Ala Pro Ser Leu Gln  
740 745 750

40 GTC ATC ACA TCC AAG CAG AGG CCC CGG AAA TTG ACA CTT ATG GGC AGC 2304  
Val Ile Thr Ser Lys Gln Arg Pro Arg Lys Leu Thr Leu MET Gly Ser  
755 760 765

45 AAC GGA CAT GAG TTT GTT TTC CTT CTA AAA GGC CAT GAA GAT CTG CGC 2352  
Asn Gly His Glu Phe Val Phe Leu Leu Lys Gly His Glu Asp Leu Arg  
770 775 780

50 CAG GAT GAG CGT GTG ATG CAG CTC TTC GGC CTG GTT AAC ACC CTT CTG 2400  
Gln Asp Glu Arg Val MET Gln Leu Phe Gly Leu Val Asn Thr Leu Leu  
785 790 795 800

45 GCC AAT GAC CCA ACA TCT CTT CGG AAA AAC CTC AGC ATC CAG AGA TAC 2448  
Ala Asn Asp Pro Thr Ser Leu Arg Lys Asn Leu Ser Ile Gln Arg Tyr  
805 810 815

50 GCT GTC ATC CCT TTA TCG ACC AAC TCG GGC CTC ATT GGC TGG GTT CCC 2496  
Ala Val Ile Pro Leu Ser Thr Asn Ser Gly Leu Ile Gly Trp Val Pro  
820 825 830

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	CAC TGT GAC ACA CTG CAC GCC CTC ATC CGG GAC TAC AGG GAG AAG AAG	2544
	His Cys Asp Thr Leu His Ala Leu Ile Arg Asp Tyr Arg Glu Lys Lys	
	835 840 845	
5	AAG ATC CTT CTC AAC ATC GAG CAT CGC ATC ATG TTG CGG ATG GCT CCG	2592
	Lys Ile Leu Leu Asn Ile Glu His Arg Ile MET Leu Arg MET Ala Pro	
	850 855 860	
10	GAC TAT GAC CAC TTG ACT CTG ATG CAG AAG GTG GAG GTG TTT GAG CAT	2640
	Asp Tyr Asp His Leu Thr Leu MET Gln Lys Val Glu Val Phe Glu His	
	865 870 875 880	
15	GCC GTC AAT AAT ACA GCT GGG GAC GAC CTG GCC AAG CTG CTG TGG CTG	2688
	Ala Val Asn Asn Thr Ala Gly Asp Asp Leu Ala Lys Leu Leu Trp Leu	
	885 890 895	
	AAA AGC CCC AGC TCC GAG GTG TGG TTT GAC CGA AGA ACC AAT TAT ACC	2736
	Lys Ser Pro Ser Ser Glu Val Trp Phe Asp Arg Arg Thr Asn Tyr Thr	
	900 905 910	
20	CGT TCT TTA GCG GTC ATG TCA ATG GTT GGG TAT ATT TTA GGC CTG GGA	2784
	Arg Ser Leu Ala Val MET Ser MET Val Gly Tyr Ile Leu Gly Leu Gly	
	915 920 925	
25	GAT AGA CAC CCA TCC AAC CTG ATG CTG GAC CGT CTG AGT GGG AAG ATC	2832
	Asp Arg His Pro Ser Asn Leu MET Leu Asp Arg Leu Ser Gly Lys Ile	
	930 935 940	
30	CTG CAC ATT GAC TTT GGG GAC TGC TTT GAG GTT GCT ATG ACC CGA GAG	2880
	Leu His Ile Asp Phe Gly Asp Cys Phe Glu Val Ala MET Thr Arg Glu	
	945 950 955 960	
35	AAG TTT CCA GAG AAG ATT CCA TTT AGA CTA ACA AGA ATG TTG ACC AAT	2928
	Lys Phe Pro Glu Lys Ile Pro Phe Arg Leu Thr Arg MET Leu Thr Asn	
	965 970 975	
	GCT ATG GAG GTT ACA GGC CTG GAT GGC AAC TAC AGA ATC ACA TGC CAC	2976
	Ala MET Glu Val Thr Gly Leu Asp Gly Asn Tyr Arg Ile Thr Cys His	
	980 985 990	
40	ACA GTG ATG GAG GTG CTG CGA GAG CAC AAG GAC AGT GTC ATG GCC GTG	3024
	Thr Val MET Glu Val Leu Arg Glu His Lys Asp Ser Val MET Ala Val	
	995 1000 1005	
45	CTG GAA GCC TTT GTC TAT GAC CCC TTG CTG AAC TGG AGG CTG ATG GAC	3072
	Leu Glu Ala Phe Val Tyr Asp Pro Leu Leu Asn Trp Arg Leu MET Asp	
	1010 1015 1020	
50	ACA AAT ACC AAA GGC AAC AAG CGA TCC CGA ACG AGG ACG GAT TCC TAC	3120
	Thr Asn Thr Lys Gly Asn Lys Arg Ser Arg Thr Arg Thr Asp Ser Tyr	
	1025 1030 1035 1040	

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TCT GCT GGC CAG TCA GTC GAA ATT TTG GAC GGT GTG GAA CTT GGA GAG 3168  
Ser Ala Gly Gln Ser Val Glu Ile Leu Asp Gly Val Glu Leu Gly Glu  
1045 1050 1055

5 CCA GCC CAT AAG AAA ACG GGG ACC ACA GTG CCA GAA TCT ATT CAT TCT 3216  
Pro Ala His Lys Lys Thr Gly Thr Thr Val Pro Glu Ser Ile His Ser  
1060 1065 1070

10 TTC ATT GGA GAC GGT TTG GTG AAA CCA GAG GCC CTA AAT AAG AAA GCT 3264  
Phe Ile Gly Asp Gly Leu Val Lys Pro Glu Ala Leu Asn Lys Lys Ala  
1075 1080 1085

15 ATC CAG ATT ATT AAC AGG GTT CGA GAT AAG CTC ACT GGT CCG GAC TTC 3312  
Ile Gln Ile Ile Asn Arg Val Arg Asp Lys Leu Thr Gly Arg Asp Phe  
1090 1095 1100

20 TCT CAT GAT GAC ACT TTG GAT GTT CCA ACG CAA GTT GAG CTG CTC ATC 3360  
Ser His Asp Asp Thr Leu Asp Val Pro Thr Gln Val Glu Leu Ile  
1105 1110 1115 1120

25 AAA CAA GCG ACA TCC CAT GAA AAC CTC TGC CAG TGC TAT ATT GGC TGG 3408  
Lys Gln Ala Thr Ser His Glu Asn Leu Cys Gln Cys Tyr Ile Gly Trp  
1125 1130 1135

25 TAC CCT TTC TGG TAA 3423  
Tyr Pro Phe Trp  
1140